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## **Biogeochemical interactions between aged cementitious materials and sulfate reducing microbial community with propionate as electron donor in the context of nuclear waste repository**

Nadège Durban, Alexandra Bertron, Vanessa Sonois-Mazars, Maud Schiettekatte, Gerald Matar, Pierre Albina, Achim Albrecht, Jean-Charles Robinet, Benjamin Erable

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1 **Biogeochemical interactions between aged cementitious materials and sulfate reducing**  
2 **microbial community with propionate as electron donor in the context of nuclear waste**  
3 **repository**

4 Nadège Durban <sup>1,2</sup>, Alexandra Bertron <sup>1\*</sup>, Vanessa Sonois-Mazars <sup>1</sup>, Maud Schiettekatte <sup>1</sup>, Gerald  
5 Matar <sup>1,2</sup>; Pierre Albina<sup>1,2</sup>, Achim Albrecht <sup>3</sup>, Jean-Charles Robinet <sup>3</sup>, Benjamin Erable <sup>2</sup>

6

7 <sup>1</sup> Laboratoire Matériaux et Durabilité des Constructions, Université de Toulouse, UPS, INSA. 135  
8 avenue de Ranguueil, 31077 Toulouse CEDEX 04, France; nadege.durban@gmail.com (N.D.);  
9 bertron@insa-toulouse.fr (A.B.); vmazars@insa-toulouse.fr (V.M.); schiette@insa-toulouse.fr (M.S);  
10 matar@insa-toulouse.fr (G.M.); pierre.albina@outlook.fr (P.A.)

11 <sup>2</sup> Laboratoire de Génie Chimique, Université de Toulouse, CNRS, INPT, UPS, 31030 Toulouse, France;  
12 benjamin.erable@ensiacet.fr (B.E.)

13 <sup>3</sup> Andra (Agence nationale pour la gestion des déchets radioactifs), 92298 Châtenay-Malabry, France;  
14 achim.albrecht@andra.fr (A.A.); jean-charles.robinet@andra.fr (J. C.R.)

15 \* Corresponding author: postal address: LMDC, INSA Toulouse, 135 avenue de Ranguueil, 31077  
16 Toulouse, Cedex 4, France. Email: bertron@insa-toulouse.fr

17 **Abstract**

18 Heterotrophic microbial sulfate reduction can occur in both natural and engineered systems. The  
19 process can influence the radionuclide speciation and mobility in deep repository of radioactive  
20 waste (DRRW). DRRW are characterised by significant masses of concrete, imposing alkaline pH in the  
21 waste cell. This paper aims to evaluate microbial sulfate reduction coupled with propionate oxidation  
22 influenced by the alkalinity of the environment: moderate alkaline pH close to 9.0 without cement  
23 paste, and moderate alkaline pH which was then moderately increased (pH 9.0 to 9.4) by the

24 presence of pre-aged solid cement pastes via advanced and moderate ageing protocols, respectively.  
25 Regardless of the degree of ageing of the cement pastes, the sulfate reduction rate decreased by up  
26 to 84% when the pH increased from 9.0 to 9.4 in the presence of cement paste and by up to 90% in  
27 the absence of cement paste. No sulfate reduction or propionate oxidation was observed for pH >  
28 9.5. Microbial metabolites, sulfide, CO<sub>2</sub> and acetate, produced from the reduction of sulfate and  
29 oxidation of propionate, and the presence of the microorganisms attached to the cement pastes  
30 (composed of up to 80% of sulfate reducing bacteria) led to their biodeterioration. Sulfide  
31 enrichment, precipitation of secondary ettringite, and intensified decalcification were notably  
32 detected. Self-healing like phenomena were also observed: calcium leached from the cementitious  
33 phases reacted with carbonate produced by microorganisms to form of calcium carbonate relocated  
34 either in the micro-cracks or on the surface of the pre-aged solid cement pastes.

35

36 **Keywords:** Sulfate-reducing bacteria; cementitious materials; nuclear waste repository;  
37 biodeterioration, moderately alkaline pH

38

## 39 **1. Introduction**

40 The management of toxic and radioactive waste often relies on clay rocks as a geological barrier and  
41 concrete as an engineered barrier to isolate the wastes from the biosphere. Many types of these  
42 wastes contain organic matter in various forms (Abrahamsen et al., 2015), some of which are known  
43 to complex both toxic metals and radionuclides present in the waste, thus increasing their solubility  
44 and mobility (Davis, 1984). For the case of radioactive waste, classified as high-level, intermediate-  
45 level or low-level radioactive waste based on its radionuclide content and activity (Ewing et al.,  
46 2016), the role of clay, as host rock, and concrete, as major engineering material, needs to be  
47 considered explicitly in the assessment of biogeochemical reactions. In France, an ongoing industrial  
48 project aiming at constructing a radioactive waste repository within Callovo-Oxfordian (COx) host  
49 rocks at a depth of 500 metres is planned for storing high-level waste and long-lived intermediate-

50 level radioactive wastes (LLILW) in concrete matrices (Albrecht et al., 2013; Rafrafi et al., 2017,  
51 2015). The stabilized LLILW will be cast in cylindrical concrete and / or steel containers, which  
52 constitute the primary packages, and will be grouped into secondary packages made of reinforced  
53 concrete (Rafrafi et al., 2017, 2015). After closing of the waste galleries, the infiltration of ground  
54 water within the storage cells is intended and is expected to induce leaching processes at the  
55 interface of cementitious and waste materials, notably. Due to these leaching phenomena,  
56 progressive enrichment of the infiltrated water is expected, such as an enrichment in cations ( $\text{Ca}^{2+}$ ,  
57  $\text{Na}^+$ ,  $\text{K}^+$ ) and hydroxide anion, which impose an alkaline pH, due to the leaching of concrete material  
58 (Bertron, 2014). An enrichment in dissolved organic matter or oxyanions released by the radioactive  
59 waste (Abrahamsen et al., 2015) is also expected. The leaching of some of these dissolved organic  
60 acids may promote the mobility of some usually immobile radionuclides by complexing them, thus  
61 affecting the safety of the repository over time, by increasing the risk of contamination of the host  
62 rock (Abrahamsen et al., 2015; Albrecht et al., 2013). Biological activity has been observed in the  
63 context of in-situ experiments in the Meuse/Haute Marne underground research laboratory in the  
64 presence of organic matter (Vinsot et al., 2008a). Therefore, a biotic oxidation of soluble organic  
65 acids can be expected in the future within the storage. Organic acids would thus be less available to  
66 form complexes with radionuclides and would therefore limit their migration into the host rock and  
67 thus improve the safety of the system. The oxidation of organic matter by microorganisms under  
68 anaerobic conditions can take place without an electron acceptor, as a fermentation process, or in  
69 the presence of an electron acceptor such as nitrate (denitrification process) or sulfate (sulfate  
70 reduction process) (Muyzer and Stams, 2008). Sulfate is an oxyanion of wider importance as an  
71 electron acceptor, given its high solubility. It may be introduced by natural water during resaturation  
72 (Vinsot et al., 2008b) and released from the waste package (bituminized waste or in resin) or cement  
73 material (Abrahamsen et al., 2015; Dauzères et al., 2014; van Loon and Hummel, 1999). With a very  
74 low standard reduction potential, i.e.  $E^0$  for sulfate/sulfite redox couple: -516 mV, the sulfate  
75 reduction is not thermodynamically favourable but it could be catalysed by sulfate-reducing bacteria

76 (SRB) under anaerobic conditions with a temperature below 60°C (Machel, 2001). SRB are versatile in  
77 terms of their metabolism and their capacities to use different sources of organic or inorganic matter  
78 (Muyzer and Stams, 2008) and the environmental conditions under which they can thrive: anaerobic  
79 wastewater treatment plants, marine and fresh water sediments, hydrocarbon seeps, hypersaline  
80 environments such as marine sediment, acid mine drainage contaminated sites, or soda lakes, where  
81 pH can be higher than 10 (Muyzer and Stams, 2008; Pallud and Van Cappellen, 2006; Sorokin et al.,  
82 2014). The metabolism of SRB is increasingly being studied for industrial applications, such as the  
83 bioproduction of metal metallic sulfide nanoparticles (such as zinc sulfide) and bioplastic  
84 (polyhydroxyalkanoate), and also for industrial wastewater treatment (Duran and Seabra, 2012; Hai  
85 et al., 2004; Qian et al., 2019; Ragu Nandhakumar et al., 2022). Very few studies have investigated  
86 the reduction of sulfate rate (SRR) at alkaline pH. Some studies have reported SRRs of the order of 18  
87 mM/d at pH 9.0 in haloalkaliphilic media (Sousa et al., 2015) or 0.023 mM/d at pH 9.5 (Mu et al.,  
88 2019). Their sensitivity to environmental physicochemical parameters, such as pH, salinity, source of  
89 organic matter and temperature, are particularly mentioned in the literature (Gutierrez et al., 2009;  
90 Muyzer and Stams, 2008; Qian et al., 2019; Sorokin et al., 2014; vanGinkel et al., 1997). However, the  
91 reduction of sulfate by SRB in a cementitious environment, and more precisely under conditions  
92 close to those found in nuclear waste repositories, has not been studied. However, it is important to  
93 evaluate the sulfate reduction as well as the oxidation rate of organic matter (i.e. propionate here)  
94 and the impact of pH on these reactions in order to reduce uncertainties in predictive models of the  
95 geochemical evolution of storage cells in underground nuclear waste repositories.

96 As mentioned earlier, in the absence of an electron acceptor, organic matter can be oxidized in the  
97 presence of micro-organisms by fermentation. Alkaline fermentation (i.e. at pH 10) produces very  
98 little methane (methanogenic process inhibited) but in comparison to neutral or slightly acidic pH  
99 values, this fermentation induces production of up to three times more VFAs (Chen et al., 2021),  
100 acetate and propionate being the main VFAs produced (Liu et al., 2014). Propionate has also been  
101 found as a degradation product of complex organic matter (such as isosaccharinic acid) identified in

102 LLIL nuclear waste (Abrahamsen et al., 2015; Kuippers et al., 2015; Rout et al., 2015). For this reason,  
103 propionate was chosen as the model organic material in this study, in order to investigate the  
104 oxidation of organic matters coupled with the sulfate reduction.

105 In this context, the objectives of the work presented in this manuscript were (i) to identify the pH  
106 limits up to which biological oxidation of propionate can be achieved via reduction of sulfate, and (ii)  
107 to measure the propionate oxidation and sulfate reduction rates in an environment like the one of  
108 the French nuclear waste repository project (moderate alkaline pH imposed by cementitious material  
109 leaching, and anaerobic conditions), and (iii) to study the possible interaction between  
110 microorganisms and cementitious materials from two perspectives. First, we observed the  
111 colonization of the material surfaces by microorganisms. And secondly, we observed the impact on  
112 the materials of the vicinity of biological activity producing sulfides, organic acids and other  
113 aggressive compounds that can promote the alteration of cementitious material (Bertron, 2014;  
114 Voegel et al., 2019b).

115 In order to assess the effect of moderately alkaline environment, the experiments were conducted at  
116 pH between 8.5 and 10. Moreover, to mimic short-term and long-term cement ageing in the nuclear  
117 waste repository (Bertron, 2014; Lagerblad and Traegaardh, 1994; Sellier et al., 2011), the cement  
118 pastes used underwent two different levels of alteration (a moderate aging (MA) and an advanced  
119 aging (AA)) where three different ratios of solid surface area to liquid volume were applied. In order  
120 to investigate biogeochemical interactions between cementitious matrices and microorganisms, the  
121 SRRs and the propionate oxidation rates (PORs) were calculated for different ratios of surface area  
122 (SA) to liquid volume (V), i.e. SA/V, and for different pH values. Indeed, increasing the SA/V ratio lead  
123 to an increase in the alkalinity of the reaction medium in terms of pH and cation concentration from  
124 the cementitious matrices leaching. Microscopic observations and complementary experiments were  
125 also performed in order to investigate the presence of microorganisms adhering to the cement paste  
126 surfaces. To evaluate the level of alteration induced by biological activity on cement paste surfaces,

127 microstructure analyses were performed and the chemical composition of the cement pastes was  
128 determined before and after their exposure to the microbial redox activity.

## 129 **2. Materials and methods**

### 130 **2.1. Preparation of cementitious material**

131 Cement pastes were prepared according to the French standard NF EN 196-3A1. Samples of  
132 CEM V/A (S-V) 42.5 N CE PM-ES-CP1 NF, from the Calcia cement factory of *Rombas* (France) were  
133 prepared with a water-to-cement ratio of 0.4 (Durban et al., 2020, 2018; Rafrafi et al., 2017) and  
134 were cast in hermetic cylindrical moulds 50 mm high and 50 mm in diameter. The cement pastes  
135 were maintained at high relative humidity (> 98 %) and 22°C for hardening during 28 days. They were  
136 then sawn into 5- or 9-mm thick slices, and each slice was sawn into eight parts to obtain coupons  
137 with a projected surface area of respectively 8 or 11 cm<sup>2</sup>. The surfaces of the final cement coupons  
138 were polished using silicon carbide polishing disks (P120 -  $\approx$  127  $\mu$ m - Presi®) to obtain a  
139 homogeneous surface roughness (Durban et al., 2020; Voegel et al., 2019b, 2019a).

140 To prepare cement leachate solutions, 2 g of crushed cement paste (granulometry 0.5 - 1.25  
141 mm) was stirred (800 rpm) into 2L of ultra-pure water deaerated with nitrogen gas in order to limit  
142 carbonation phenomena between leached calcium and CO<sub>2</sub> solubilized in the water. The leaching  
143 process maintained anaerobically was stopped when the water solution conductivity reached  
144 between 500 and 550  $\mu$ S/cm with a pH above 11.0. Finally, the cement leachate was filtered and  
145 then stored at 4°C in hermetically sealed bottles saturated by nitrogen gas or used immediately  
146 (Albina et al., 2021). The chemical composition of the cement leachate solution was analysed using  
147 Inductively Coupled Plasma-Optical Emission Spectrometry (ICP-OES) and the results are reported in  
148 Table S.1.

### 149 **2.2. Ageing processes for cement pastes**

150 A typical cement leachate from freshly poured cement paste has an alkaline pH between 12 and 13.  
151 This high pH value decreases with time due to various ageing processes that enhance leaching of  
152 hydroxide ions and cations such as calcium ions (Bary, 2008; Bertron, 2014; Gallé et al., 2006; Planel

153 et al., 2006). Two ageing protocols were set up in order to mimic short-term and long-term cement  
154 ageing in the nuclear waste repository.

### 155 *2.2.1 Advanced ageing process*

156 Cement paste coupons with a thickness of 9 mm (projected surface area 11 cm<sup>2</sup>) were firstly  
157 immersed in bottles containing 1 L of 0.28 mM acetic acid solution supplemented with 1 mM NaOH  
158 to reach pH 4 for three days (solid surface area to liquid volume ratio 64 cm<sup>2</sup>/L). The solution was  
159 renewed as soon as the pH was higher than 4.3 (Bertron et al., 2007; Larreur-Cayol et al., 2011).  
160 Secondly, the coupons were immersed in ultra-pure water for seven days with 3 renewals of the  
161 solution in order to remove all traces of acetic acid. During the ageing process, the bottles were  
162 continuously stirred (800 rpm), at room temperature, and N<sub>2</sub> bubbling was applied after each  
163 solution renewal in order to avoid carbonation phenomena. Alteration of cementitious materials by  
164 acetic acid has been thoroughly investigated, and this technique allows fast leaching of hydroxide  
165 (OH<sup>-</sup>) and alkaline cations (Ca<sup>2+</sup>, Na<sup>+</sup>, K<sup>+</sup>) without promoting any secondary precipitation or any  
166 specific interactions between the acetate anion (CH<sub>3</sub>COO<sup>-</sup>) and the cations leached from cement  
167 pastes (Bertron et al., 2007, 2005a, 2005b; Bertron and Duchesne, 2013). The cement samples  
168 treated in this way were labelled AA (advanced ageing).

### 169 *2.2.2 Moderate ageing process*

170 Cement paste coupons were immersed in 2 L (solid surface area to liquid volume ratio 43 cm<sup>2</sup>/L) of  
171 ultra-pure water at room temperature for 8 days, under agitation (800 rpm) and N<sub>2</sub> bubbling, the  
172 water being renewed on a daily basis. Compared to the advanced ageing process, the moderate  
173 ageing process creates less damage to the surface of cement pastes (Duchesne and Bertron, 2013).  
174 The stock of cementitious cations (alkalis) (Ca<sup>2+</sup>, Al<sup>3+</sup>, Na<sup>+</sup>, K<sup>+</sup>) and hydroxide (OH<sup>-</sup>) ion in the  
175 moderately aged cement coupons, and in particular in the surface layer, was thus higher than that in  
176 the AA cement paste coupons. The moderately aged cement pastes were labelled MA (moderate  
177 ageing).

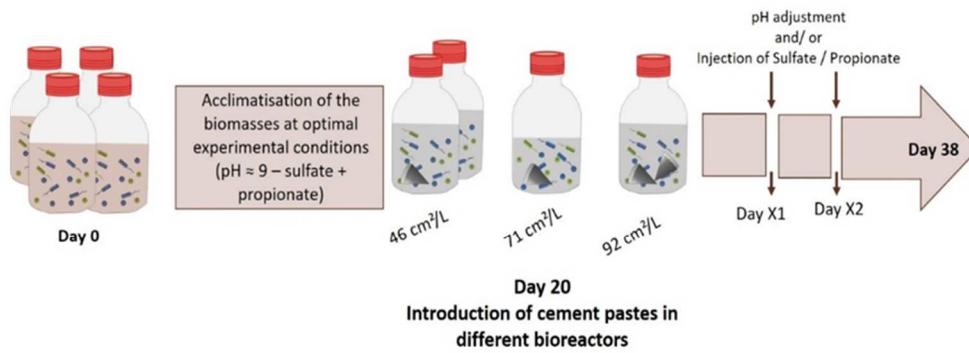
178 **2.3. Inoculum for the bioreactors**

179 The biological inoculum was a microbial consortium collected from activated sludge of the  
180 wastewater treatment plant in Castanet, France. In order to concentrate the inoculum and remove  
181 soluble organic substrates from the sludge, 250 mL of the collected mixed liquor was centrifuged at  
182 4600 g at 6°C for 15 min. Finally, the bioreactors were inoculated with 2.8 g/L of dewatered sludge.

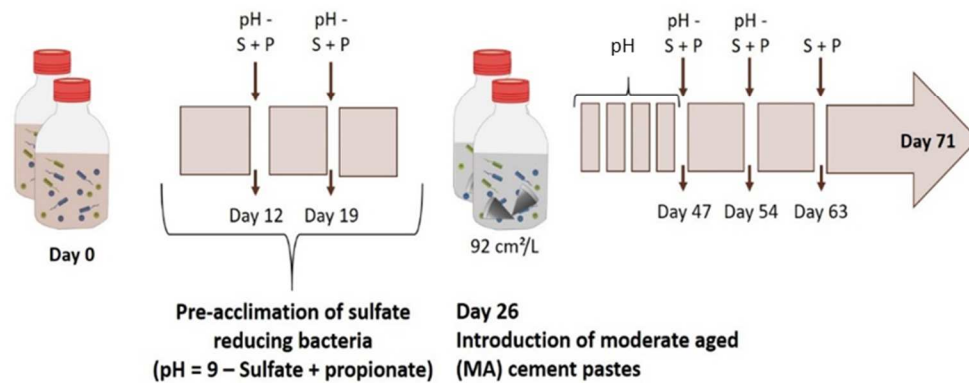
183 **2.4. Experiments in bioreactors**

184 250 mL sterile glass bottles having a rubber septum and a screw cap with an aperture were used as  
185 anaerobic bioreactors. The bioreactors were protected from light and operated in fed-batch, in  
186 anaerobic conditions and at 30 °C. Biological propionate oxidation coupled with sulfate reduction  
187 was investigated first in the cement leachate and then in the presence of AA or MA cement paste  
188 coupons. The experiments were organized in three successive steps: [step 1] a period of  
189 acclimatization for the biological inoculum, with a progressive pH increase from 8.0 to 9.0 in the  
190 cement leachate without cementitious material; [step 2] evaluation of the propionate oxidation rate  
191 (POR) and the sulfate reduction rate (SRR) without cementitious material; and [step 3] introduction  
192 of cement paste coupons, after 20 days for AA and 26 days for MA cement pastes coupons. Before  
193 starting each step, the liquid phase of the bioreactor was deaerated by N<sub>2</sub> gas bubbling for 15 min.  
194 During all the steps, regular supplementation with 5.7 mM of neutralized propionate and 10 mM of  
195 disodium sulfate was performed (Figure 1). At the end of the experiment, to confirm the presence of  
196 bacteria attached to the cement paste surface and to characterised the reactivity of microorganism  
197 attached, the protocol described by Durban et al. (2020) was adapted and applied (see section S.1 in  
198 supplementary material). The bacterial community was also investigated according to the protocol  
199 described by Durban et al. (2020) (Section S.2 in supplementary material).

**A : Experimental setup of the bioreactors in presence of AA cement pastes**



**B : Experimental setup of the bioreactors in presence of MA cement pastes**



200

201 **Figure 1:** Experimental set-up of the bioreactors to study propionate oxidation and sulfate reduction  
 202 in presence of A) advanced aged (AA) cement paste coupons and B) moderate aged (MA) cement  
 203 paste coupons. The pH was adjusted throughout the experiment, while propionate and sulfate were  
 204 added depending on the reactors as represented: pH: pH adjustment, S: Sulfate supplementation, P:  
 205 Propionate supplementation.

206

207 **2.4.1 Experiments in the presence of AA cement paste coupons**

208 A first set of experiments was carried out using four bioreactors containing inoculated cement  
 209 leachate supplemented with 5.7 mM of propionate neutralized with NaOH (pH: 7.5) and 10 mM of  
 210 disodium sulfate. The pH was initially adjusted to 8 and was progressively increased with NaOH 1M  
 211 addition to reach 9, i.e. a moderately alkaline condition. During [step 2], after sulfate and propionate  
 212 supplementation, the liquid phases in the bioreactors (see section 2.5) were sampled to evaluate the  
 213 POR and the SRR. For [step 3], AA cement paste coupons were added at increasing surface to  
 214 volume ratios (area of cement paste surface/cement leachate volume: 46, 71 and 92 cm<sup>2</sup>/L), in order  
 215 to increase the effect of the cementitious material on microbial activity (Figure 1.A). The pH was  
 216 maintained at values between 9 and 9.5 using hydrochloric acid (1M).

#### 217 *2.4.2 Experiments in the presence of MA cement pastes*

218 A second series of experiments was carried out using two bioreactors supplemented with 5.7 mM of  
219 propionate neutralized with NaOH (pH: 7.5) and 10 mM of disodium sulfate. The initial pH was  
220 adjusted to 8.8 and the PORs and SRRs were evaluated [step 2]. For [step 3], the MA cement paste  
221 coupons were added at day 26 (Figure 1.B) with a surface to volume ratio of 92 cm<sup>2</sup>/L. The pH was  
222 maintained between 10 and 9.0 for 16 days by using hydrochloric acid (1M). Once the pH was  
223 stabilized, the liquid phase in bioreactors (see section 2.5) was regularly sampled to evaluate the  
224 PORs and the SRRs in the presence of cement paste.

#### 225 **2.5. Chemical analysis of the inoculated cement leachate**

226 For the samples collected with sterile syringes from the liquid fraction of the bioreactors, i.e. the  
227 inoculated cement leachate, the pH was immediately measured using a Eutech Instruments 6500 pH/  
228 ion meter, and to monitor microbial growth, the optical density was measured at 600 nm using a  
229 JENWAY 7315 spectrophotometer (Rafrafi et al., 2015).

230 2 mL of collected solutions were also filtered using 0.2 µm filters (196 Minisart® PES, Sartorius) for  
231 analyses of ionic species. The concentrations of propionate, acetate, sulfate, sulfite and thiosulfate  
232 were quantified by High Performance Ion Chromatography (Dionex ICS-2000 and ICS-3000) according  
233 the protocol described by Alquier et al. (2014) modified according the KOH gradient described in  
234 Table S.2.

#### 235 **2.6. Analyses of solid cementitious materials**

##### 236 *2.6.1 Microstructural observation and chemical analyses with scanning electron microscopy (SEM)* 237 *coupled to energy dispersive X-ray spectroscopy (EDS)*

238 The microstructural and chemical characteristics of the aged cement coupons before and after their  
239 exposure in the bioreactor were analysed by SEM- EDS (JEOL 6380-LV coupled with XFlash 6/30  
240 Bruker EDS detector). The cement paste coupons were sawn perpendicularly to their flat surfaces.  
241 One portion of the specimen was embedded in epoxy resin (Mecaprex Ma2+, Presi). The sawn  
242 surface comprising the outer zone and the unaltered core of the specimen was dry polished with  
243 silicon carbide polishing disks (Presi) (polishing disc references and grain sizes: ESCIL, P800–22 µm,

244 P1200–15 µm and P4000–5 µm). The polished sections were then coated with carbon (Giroudon et  
245 al., 2021; Voegel et al., 2019a).

#### 246 *2.6.2 Mineralogical analyses with X-ray diffraction (XRD)*

247 The mineralogical composition of the cement paste coupons was analysed before and after their  
248 exposure in the different bioreactors using X-ray diffraction (BRUKER D8 Advance, Cu cathode, 40 kV,  
249 40 mA). An initial analysis was carried out on the external surface of the coupons that was directly  
250 exposed to the liquid fraction of the bioreactors. Then the external layer was cautiously scraped  
251 using silicon carbide polishing disks (P120 -  $\approx$  127 µm - Presi®), until a depth of 100-200 µm was  
252 achieved. This procedure was repeated several times to remove the precipitates from the surfaces of  
253 cement pastes, if any, and to eliminate the altered zones until unaffected core was reached within  
254 the cement coupons (Bertron et al., 2009).

#### 255 **2.7. Fluorescence microscopy**

256 Biofilms attached and developed on the cement paste coupons were chemically fixed following a  
257 specific biofilm fixation protocol on cement pastes, as described previously (Durban et al., 2020;  
258 Voegel et al., 2015). The labelling step consisted of submerging the coupons in a mixture of PBS  
259 (phosphate buffered saline) solution and SYTO® 9 (Invitrogen, USA) for 10 minutes. The final step  
260 before microscopic visualization consisted of rinsing the cement coupons with 10 mM PBS solution  
261 (phosphate buffered saline - Commercial tablets pH 7.2, Applichem), to remove excess dye (Durban  
262 et al., 2020). Sample visualization was performed using a Carl Zeiss AxioImager M2 microscope  
263 designed for epifluorescence detection and equipped with an HBO 200 W/4 and Zeiss set 09 as a  
264 filter (excitor HP450-490, reflector FT 10, barrier filter LP520). Images were acquired using a  
265 monochrome digital camera, and several acquisitions (about 70 images) were performed and  
266 compiled using the Zen blue software. To avoid potential rapid carbonation of the cement coupon  
267 during the imaging, the cement coupons were immersed in a 10X PBS solution.

### 268 **3. Results**

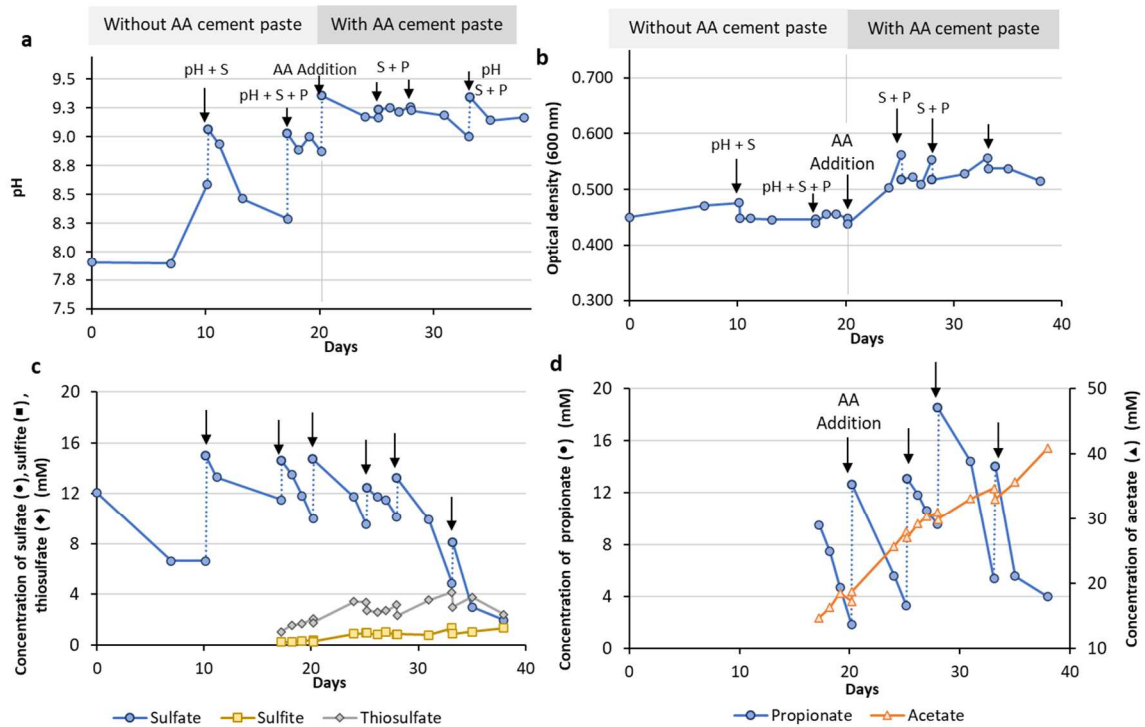
269 Reactors under abiotic conditions were operated to evaluate the reactivity of the systems in the  
270 absence of microorganisms. No uptake was observed for propionate, a low uptake of sulfate (0.02 -

271 0.04 mM/d) was observed (result are availed as supplementary data Figure S.3). A white needle-like  
272 deposit (ettringite) was observed on the surface of the cement pastes in SEM (data not shown).

### 273 **3.1 Assessment of the microbial propionate oxidation and sulfate reduction in the** 274 **presence of AA cement paste coupons**

#### 275 *3.1.1 Analysis of the liquid fraction of the bioreactors*

276 The SRR and POR were investigated in the presence and absence of AA cement paste coupons with  
277 different solid surface area to liquid volume ratios (SA/V ratios): the two duplicates for 46 cm<sup>2</sup>/L, and  
278 71 and 92 cm<sup>2</sup>/L. The results were very similar for the four bioreactors. Consequently, only the  
279 results of the bioreactor containing the SA/V ratio of 71 cm<sup>2</sup>/L is discussed below and the results of  
280 the other ratios studied are reported in the supplementary data (Figure S.4, Figure S.5 and Figure  
281 S.6). Figure 2 thus shows the evolution of the pH (Figure 2.a) and the optical density (OD) at 600 nm  
282 (Figure 2.b) in the bioreactor containing the SA/V ratio of 71 cm<sup>2</sup>/L, and the evolution of  
283 concentrations of sulfate, sulfite, and thiosulfate (Figure 2.c), and propionate and acetate (Figure  
284 2.d), between day 17 and the end of the experiment. AA cement paste coupons were incorporated  
285 into the bioreactor at day 20. The data collected between day 0 and day 17 cover the period of  
286 acclimation to the pH increase from 8 to 9 (step 1) and are not of interest for the current study.



287

288 **Figure 2:** Evolution of the pH (a) and the Optical Density OD (b) within the bioreactors into which AA  
 289 cement pastes coupons were incorporated on day 20 with a solid surface to liquid volume ratio of 71  
 290 cm<sup>2</sup>/L. Variations in concentrations of sulfate, sulfite and thiosulfate (c), and propionate and acetate  
 291 (d). The black arrows indicate incorporation of cementitious materials, sulfate (S) and propionate (P)  
 292 additions, and pH adjustments.

293

294 During step 2, between days 17 and 20, the average OD was 0.450 in the bioreactor with an SA/V  
 295 ratio of 71 cm<sup>2</sup>/L. The sulfate was reduced to sulfite and thiosulfate, which reached the maximal  
 296 concentrations of 0.4 mM and 2 mM, respectively, on day 20. Approximately 70% of the carbon from  
 297 the propionate oxidation was converted into acetate and CO<sub>2</sub>. Over all the bioreactors, the average  
 298 SRR was 1.42 ± 0.13 mM/d and the average POR was 1.66 ± 0.58 mM/d, with an average pH of 9.1  
 299 (Table 2)

300 Immediately after the introduction of the AA cement paste coupons, an increase in pH from 8.9 to  
 301 9.4 was observed on day 20 (Figure 2.a). The same phenomenon was observed on day 33  
 302 immediately after bubbling. The OD increased steadily from 0.45 to 0.52 between days 20 and 26,  
 303 probably due to precipitates formed in solution from the reaction between cementitious ions  
 304 released by the AA cement paste coupons, then the OD remained constant between day 26 and day

305 39. The presence of solid cement pastes did not affect the OD significantly (Figure 2.b). Sulfate was  
306 reduced into thiosulfate and sulfite. Their concentrations increased and then stabilized after about 8  
307 days to remain close to 0.9 mM of sulfite and 4.0 mM of thiosulfate (Figure 2.c). Figure 2.d shows  
308 propionate removal and the simultaneous formation of acetate that accumulated in the bioreactor;  
309 71% of the carbon from the oxidation of propionate was used to form acetate.

310

311 Table 1 summarizes the rates of sulfate reduction and propionate oxidation for the bioreactors  
312 operated with different values of SA/V, before and after the addition of AA cement paste coupons.  
313 Regarding the effect of the SA/V ratio at pH 9.2, the SRRs corresponding to SA/V ratios of 46 and 71  
314 cm<sup>2</sup>/L, 1.44 and 1.54 mM/d, respectively. These ratio values were similar to the SRR obtained  
315 without AA cement paste coupons, 1.40 cm<sup>2</sup>/L at pH 9.1 (Table 1). Nonetheless, the SRR decreased to  
316 the value of 0.9 mM/d in the bioreactor containing the SA/V ratio of 96 cm<sup>2</sup>/L. The POR was also little  
317 affected by SA/V ratios of 46 and 71 cm<sup>2</sup>/L – with a rate close to 2 mM/d, similar to that without  
318 cement paste – but the POR decreased to 1.4 mM/d for the reactor corresponding to the ratio of 96  
319 cm<sup>2</sup>/L. In order to evaluate the impact of pH on SRR and POR, the data obtained from the reactors  
320 with 46 cm<sup>2</sup>/L as SA/V ratios were collected and the average SRR and POR were calculated for  
321 different pH values. This highlighted a decrease of the SRR for pH above 9.4: from 1.10 mM/d at pH  
322 9.0 to 0.67 mM/d at pH 9.4, and also showed that the POR varied from 1.48 mM/d at pH 9.0 to 0.84  
323 mM/d at pH 9.4.

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333 **Table 1:** Sulfate reduction rate and propionate oxidation rate with and without AA cement paste  
 334 coupons according to different SA/V ratios or according to different pH values.

|  | Solid/liquid ratio    | Average initial pH* | POR (mM/d)  | Acetate production rate (mM/d) | SRR (mM/d)  |
|--|-----------------------|---------------------|-------------|--------------------------------|-------------|
| Without AA cement paste coupons - all reactors included (+ sd)                             |                       | 9.1 ± 0.0           | 2.10 ± 0.40 | 2.10 ± 0.90                    | 1.40 ± 0.10 |
| With AA cement paste coupons according to different solid/liquid ratios for the same pH ** | 46 cm <sup>2</sup> /L |                     | 2.04        | 1.86                           | 1.44        |
|  | 71 cm <sup>2</sup> /L | 9.2                 | 1.92        | 1.84                           | 1.54        |
|  | 92 cm <sup>2</sup> /L |                     | 1.38        | 1.38                           | 0.90        |
| With AA cement paste coupons according to different pH values for the same SA/V (+ sd)     | 46 cm <sup>2</sup> /L | 9.0 ± 0.1           | 1.48 ± 0.70 | 1.10 ± 0.20                    | 1.42 ± 0.60 |
|  |                       | 9.2                 | 1.65        | 1.10                           | 1.86        |
|  |                       | 9.4 ± 0.1           | 0.84 ± 0.60 | 0.67 ± 0.30                    | 0.82 ± 0.30 |

335

336 \* pH measured after supplementation and N<sub>2</sub> bubbling; \*\* standard deviation (sd) when the calculation is possible i.e.

337 when there were more than three measuring points \*\*\* SRR: sulfate reduction rate; POR: propionate oxidation rate

338 **3.1.2 Biological investigations**

339 The investigation of microorganisms adsorbed onto, bound by or colonizing the surfaces of AA  
 340 cement paste coupons was conducted by specific fluorescent labelling capable of revealing the  
 341 presence of microbial cells by observation with epifluorescence optical microscopy (Figure S.7 in  
 342 supplementary material). The cement paste surface was covered with rounded clumps, whose green  
 343 fluorescence suggested that they were microbial clusters about 30 μm thick. Other filamentous  
 344 structures, possibly corresponding to chains of microorganisms such as filamentous bacteria (Figure  
 345 S.7, white rectangles), were also highlighted. These structures were quite distinct from calcite  
 346 precipitates, which are sometimes capable of emitting auto-fluorescence under epifluorescence light  
 347 microscopy (Chang et al., 2003; Durban et al., 2020).

348

349 At the end of step 3, in order to confirm the presence of microorganisms attached to the cement  
 350 paste surface and to study their metabolic activity, the cement paste from one of the bioreactor with  
 351 an SA/V ratio of 46 cm<sup>2</sup>/L was transferred to a new bioreactor containing fresh cement leachate and

352 supplemented with sulfate and propionate according to the protocol described by Durban et al.  
353 (2020) (detailed protocol in supplementary data; section 1). A progressive increase of the OD (Figure  
354 S.8 in supplementary data) together with a reduction of sulfate and an oxidation of propionate  
355 (Figure S.8) confirmed that micro-organisms were indeed attached to the surface of the cement  
356 paste, because they were the only possible source of microorganisms in the bioreactor. The  
357 oxidation of propionate coupled with the reduction of sulfate increased with the recolonisation of  
358 the supernatant. Between days 8 and 15, a POR of 0.46 mM/d and SRR of 0.64 mM/d were reached  
359 while between days 15 and 21, a POR of 2.35 mM/d and SRR of 1.65 mM/d were was obtained.

360 The microbial populations that constituted the inoculum, i.e. activated sludge from domestic  
361 wastewater treatment plants, was similar to that described by Durban et al. (2020), i.e. the main  
362 phyla within the microbial community were *Proteobacteria* ( $\approx 20\%$ ), *Actinobacteria* (10%),  
363 *Planctomycetes* (9%), *Bacteroidetes* (5%), *Chloroflexi* (4%) and *Firmicutes* (3%) (Figure S.9 in  
364 supplementary material). A microbiological examination of the microbial community established  
365 close to the surface of the cement paste showed a dominance in the community of *Proteobacteria*  
366 (82%) and, more especially, of *Desulfomicrobium sp.* (80%). *Firmicutes* (2%) and *Spirochaetes* (2%)  
367 were present in lesser degrees.

### 368 **3.1.3 Analysis of cement paste coupons**

#### 369 3.1.3.1 Macroscopic observations

370

371 The ageing process induced a dense cracking pattern on the surface of AA cement paste coupons  
372 (Figure S.11.a and b), probably due to chemical shrinkage following the intense leaching of calcium  
373 and other cementitious cations (creation of porosity) (Bertron et al., 2007). The cracks were still  
374 visible on the cement pastes after their 18 days' immersion in the bioreactor at pH 9.0. Moreover,  
375 the outer layer of the cement paste turned black, likely as a result of the formation of iron sulfide  
376 resulting from the reaction between iron leached from cementitious material and sulfide formed by  
377 the reduction of sulfate by SRB (Figure S.11.c and d).

378

379 3.1.3.2 Chemical and microstructural characterization of AA cement pastes coupons before and after  
380 their exposure in the bioreactors

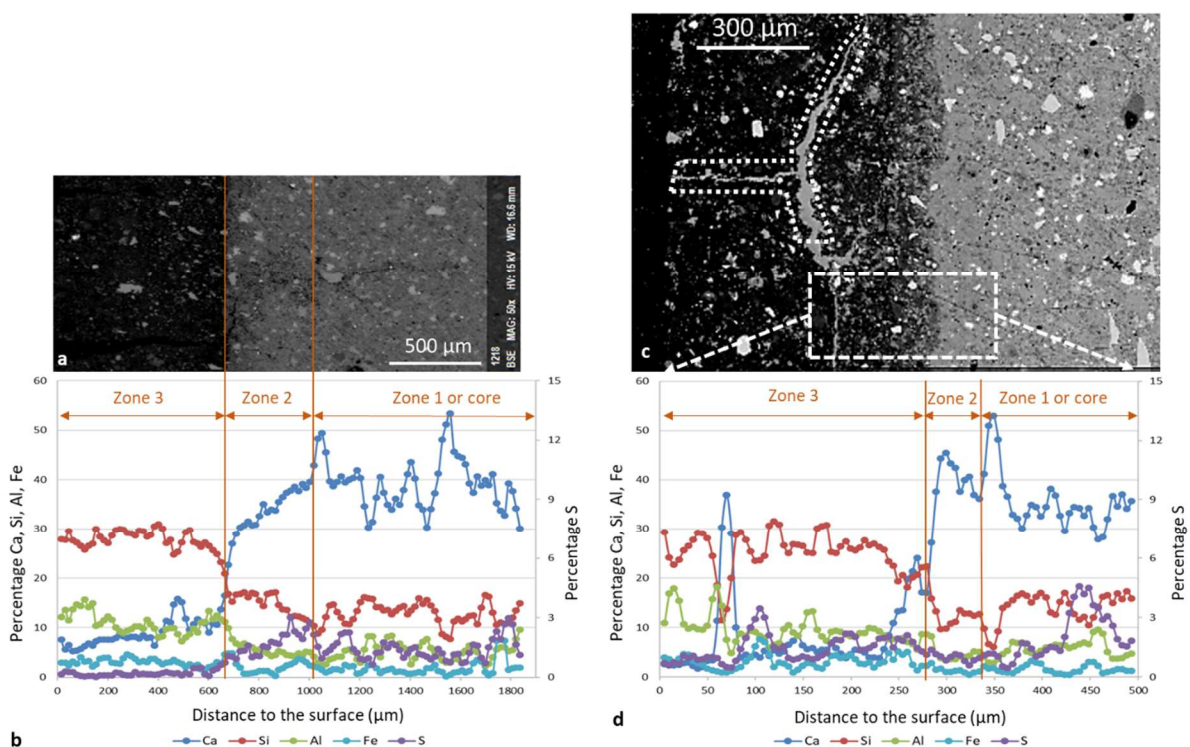
381

382 • **AA cement paste coupons before exposure:**

383 Figure 3 presents the SEM observation of an AA cement paste coupons after the pre-treatment with

384 acetic acid and ultra-pure water (Figure 3.a) and its chemical composition profile (Figure 3.b)

385 obtained with EDS.



386

387 **Figure 3:** Scanning electron micrograph (SEM) of the external micrometres of a cross section of the  
388 AA cement paste coupon before (a) and after its exposure to the culture medium in the bioreactor  
389 with a ratio of 92 cm<sup>2</sup>/L (c). Evolution of the chemical composition of the corresponding zones,  
390 mainly Ca, Si, Al, Fe, and S, according to the distance to the surface (b and d). Three different zones  
391 can be identified within the cross-section AA of the cement paste coupon: the core of the cement  
392 paste corresponds to zone 1 while the surface that was exposed to the bioreactor corresponds to  
393 zone 3. The area delimited in the upper left corner of the SEM image (b) corresponds to self-healing  
394 of the AA cement paste coupon, due to biological activity and calcite formation.

395 A chemical and microstructural zonation is clearly visible (the greyscale in SEM in backscattered  
396 electron mode depends on the intensity of the signal, and expresses the mass density of the matter).

397 Zone 1, or the sound zone/core of specimen, has the typical composition of an unaltered

398 cementitious specimen. It is mainly composed of Ca (37.4 % on average), Si (14.6 %), Al (6.1%), Fe  
399 (1.7%), and S (1.4%). It shows lighter grey residual anhydrous grains, embedded in a darker grey  
400 hydrated cement matrix. Actually, the hydration of cementitious materials reduces the average  
401 atomic number of the phases because of the water molecules integrated into the structure, and the  
402 black zones correspond to porosity. Zone 2, which can be considered as a transition zone between  
403 zones 1 and 3, is characterized by a progressive decrease of the Ca and S contents, together with a  
404 relative increase of Si, Al and Fe contents. Residual anhydrous grains are still visible in this zone. The  
405 limit between zone 2 and zone 3 is marked by a sharp decrease in the calcium content. Zone 3, the  
406 outer layer exposed to the acid solution, is darker than zones 1 and 2, indicating a lower density of  
407 the paste. This zone is indeed strongly decalcified (average Ca content 9.1 %) and S is completely  
408 leached, whereas Si, Al and Fe have been preserved, as these elements were stable during the acid  
409 attack carried out during the pre-treatment. The thicknesses of zones 2 and 3 are 670  $\mu\text{m}$  and 340  
410  $\mu\text{m}$ , respectively, and the total altered layer thickness (zone 2 + zone 3) is 1010  $\mu\text{m}$ .

411 • ***AA cement paste coupon after exposure***

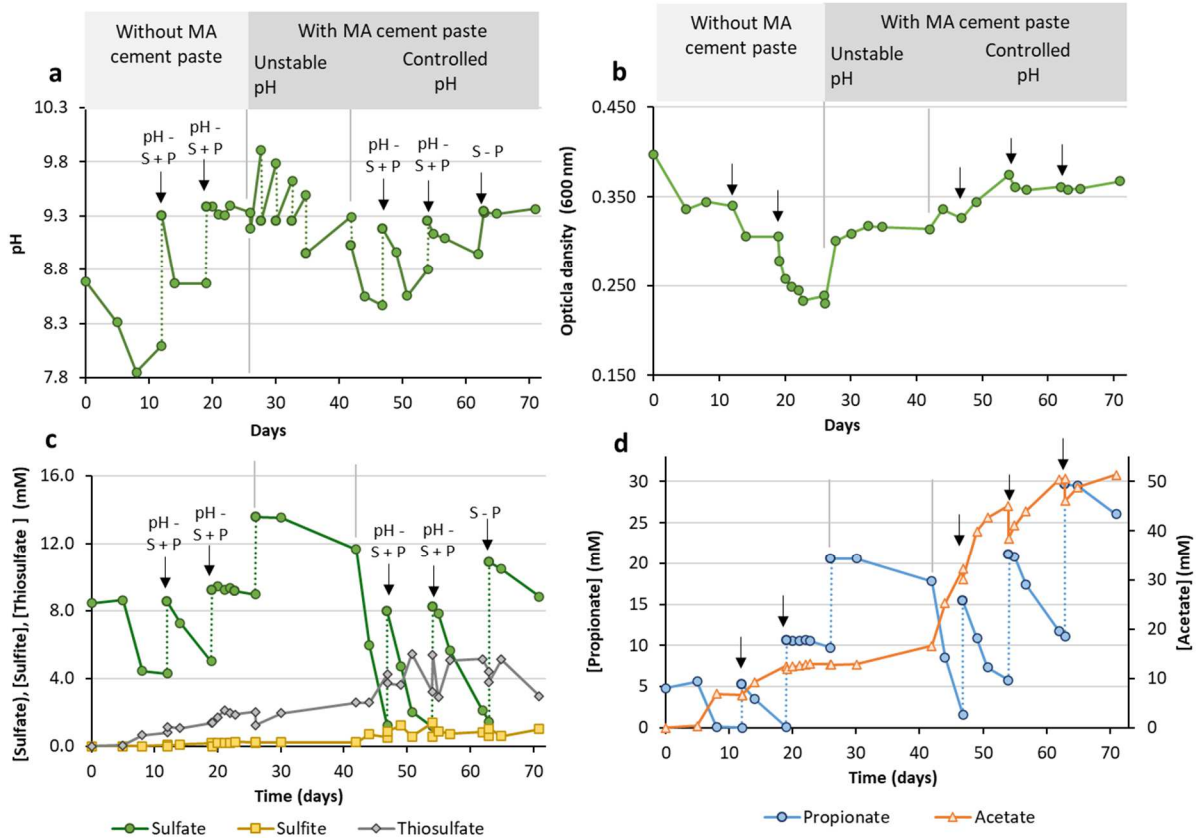
412 Three main zones can be distinguished on the Figure 3.d according to the distance to the surface in  
413 contact with the liquid medium. Zone 1 corresponds to the unaltered core of the specimen, while  
414 zone 3 corresponds to the outer layer of the cement paste, including the surface exposed to the  
415 microbial communities (Figure 3.c). Zone 1 has the typical microstructural, chemical and  
416 mineralogical pattern of a sound hydrated cement paste. Zone 2, about 83  $\mu\text{m}$  thick, is slightly  
417 decalcified, showing a much lower density of anhydrous grains and no enrichment of sulfate (Figure  
418 3.c and d). In zone 3, which is 750  $\mu\text{m}$  thick, there is a progressive, but very strong, decrease of the  
419 calcium content tending to zero in the outer layer (Figure 3.d). It is mainly composed of Si, Al, and Fe  
420 (Figure 3.d). It is also noticeable that sulfur is still present in this zone although it was absent from  
421 specimens exposed to pH 4 acetic acid solution (Figure 3.a and b). This may be due to an enrichment  
422 of exogenous sulfur diffusing from the liquid phase of the bioreactor and precipitating in the  
423 cementitious matrix. Additionally, a crack network filled with secondary precipitates is clearly visible.

424 Chemical analyses showed that these precipitates were composed of calcium, suggesting the  
425 formation of  $\text{CaCO}_3$ . This secondary precipitate is highlighted on the chemical composition profile: at  
426 280  $\mu\text{m}$ , a localized, strong increase in Ca is observed. This precipitation of calcite, clogging the  
427 cementitious matrix cracks, could be attributed to the microbial activity of SRB (Figure 3.c).  
428 SEM imaging of the surface of cement coupons highlights the presence of sphere-shaped compounds  
429 (Figure S.10), a few micrometres in diameter, within some fractured zones of the AA cement pastes  
430 coupons (Figure S.10.a - b). The results of EDS analyses show that these spheres are composed  
431 mainly of calcium (Table S.3, spectra 1 (Figure S.10.a), 4 and 6 (Figure S.10.b)). According to the X-ray  
432 diffraction analyses (Figure S.12), these compounds on the surface of the materials were calcium  
433 carbonate precipitates ( $\text{CaCO}_3$ ).  
434 Other punctual chemical analyses on the surface of the cement coupons (Table S.3, spectra 3, 5, 7)  
435 highlighted that the matrix was mainly composed of silicon, aluminium and iron, confirming the  
436 strong decalcification of the matrix (because of the AA cement paste coupon pre-treatment). The  
437 sodium content was also noticeable – and unusual considering the pre-treatment. It may have been  
438 due to an enrichment by exogenous sodium (sulfate was introduced in the bioreactor in the form of  
439 sodium sulfate and propionate was neutralised with NaOH). This type of enrichment by exogenous  
440 sodium in the outer layer of decalcified cement paste has already been reported in the literature  
441 (Bertron et al., 2005a).

### 442 **3.2 Assessment of the microbial propionate oxidation and sulfate reduction in the** 443 **presence of moderate aged (MA) cement paste coupons**

#### 444 *3.2.1 Analysis of the liquid fraction of the bioreactors*

445 The SRR and POR were investigated in duplicate in the presence and absence of MA cement paste  
446 coupon for one SA/V ratio, 92  $\text{cm}^2/\text{L}$ . Figure 4 presents the evolution of pH (a), OD (b), concentrations  
447 of sulfate, sulfite and thiosulfate (c), and the concentrations of propionate and acetate (d) in the  
448 liquid phase for one of the duplicates of the experiment (Reactor A). The results of the duplicate, the  
449 bioreactor B, are available as supplementary data (Figure S.13).



450

451 **Figure 4:** Evolution of pH (a), Optical Density (b), concentration of sulfate, sulfite, thiosulfate (c),  
 452 propionate and acetate (d) in the bioreactor into which MA cement paste coupons were inserted at  
 453 day 26, with a ratio of 92 cm<sup>2</sup>/L. The black arrows indicate pH adjustment, and sulfate (S) and  
 454 propionate (P) additions.

455 Three different steps can be distinguished, (i) the initial acclimation phase without solid cement  
 456 paste and, jointly, evaluation of the PORs and the SRRs for several pH values (steps 1 and 2: days 0 to  
 457 26), (ii) the acclimation of SRB to the chemical conditions imposed by the MA cement pastes coupons  
 458 added on day 26 (step 3a: days 26 to 42), and (iii) propionate oxidation coupled with sulfate  
 459 reduction after SRB acclimation (Step 3b: days 42 to 71) (Figure 4).

460 During step 1, the pH decreased from 8.7 to 7.9 between days 0 and 8 (Figure 4.a) while sulfate  
 461 reduction and propionate oxidation were only observed from day 5. Sulfate concentration decreased  
 462 from 8.6 mM to 4.4 mM; it was reduced into sulfite (maximal concentration of 0.2 mM) and  
 463 thiosulfate (maximal concentration of 2 mM). These reductions of the sulfate and the products from  
 464 the sulfate reduction were coupled with the oxidation of propionate into CO<sub>2</sub> and acetate, which  
 465 gradually accumulated in the bioreactor. On average, 76% of the carbon from the oxidation of

466 propionate led to the generation of acetate. A decrease in OD from 0.40 to 0.24 was observed  
467 between days 0 and 26 (Figure 4.b), due to the acclimation of microorganisms from activated sludge  
468 to the operating conditions in the bioreactor. Despite this, the biological propionate oxidation and  
469 the sulfate reduction was well established by day 12. Table 2 summarizes the SRR and POR for both  
470 bioreactors, before and after the addition of MA cement paste coupons. Before MA cement pastes  
471 coupons addition, at an average initial pH of 8.7 and 9.4, the SRR and POR decreased with the pH  
472 increase from 0.9 mM /d to 0.1 mM /d for SRR and from 1.3 mM/d to 0.1 mM/d for POR. The MA  
473 cement paste coupon addition induced an increase in pH to values around pH 10.0 on day 28 (Figure  
474 4.a). This was due to the release of hydroxide ion from MA cement paste coupons. An increase of OD  
475 values from 0.23 to 0.30 also occurred, due the formation of precipitates resulting from the leaching  
476 of cement cations (Figure 4.b). Consequently, several pH balance corrections to pH 9.0 were  
477 performed during step 3a (days 26 to 42) to avoid excessive drift towards very alkaline pH levels  
478 detrimental to the viability of microorganisms. The pH then fluctuated between 8.8 (lowest value)  
479 and 9.9 (highest value). During step 3a and after day 28, the OD remained constant with an average  
480 value of 0.31. (Figure 4.a). Only a very slight reduction in the sulfate concentration was observed  
481 during this step with an SRR of 0.2 mM/d as well as a POR of 0.2 mM/d (Figure 4.a).

482 The OD increased slightly from 0.31 on day 42 to 0.35 on day 55 and remained constant thereafter  
483 (Figure 4.b). Associated with this increase in OD and as a consequence of the pH adjustment, the  
484 beginning of phase 3b was characterized by a decrease in pH over 5 days (from 9.0 to 8.5) associated  
485 with a significant reduction of sulfate (SRR of 2.21 mM/d) coupled with an oxidation of propionate  
486 (POR of 3.46 mM/d). This step (3b) corresponded to a stabilization of the pH and a significant  
487 biological activity of sulfate reduction and propionate oxidation, which followed a latent step (step  
488 3a) where the instability of the pH was not favourable to such biological activity. This latent step  
489 lasted 16 days for the reactor A (Figure 4) and 28 days for the duplicate, reactor B (Figure S.13).

490 As previously observed, the sulfate reduction led to the formation of reactive intermediates that did  
491 not accumulate, i.e., sulfite and thiosulfate (Figure 4.c). The concentrations of sulfite and thiosulfate

492 reached were a little higher than in step 2 but of the same order of magnitude as for the experiments  
 493 with AA cement paste coupon, i.e. an average  $0.8 \pm 0.3$  mM for sulfite and  $4.1 \pm 1.0$  mM for  
 494 thiosulfate. On the other hand, acetate gradually accumulated in the system, resulting from the  
 495 oxidation of propionate. 86% of the carbon from the oxidation of propionate was used to form  
 496 acetate.

497 Table 2 reports the SRRs and PORs calculated for several pH values before the addition of MA  
 498 cement pastes coupons and after their addition into the bioreactor at day 26. Before cement paste  
 499 addition, the PORs decreased as the pH increased from 1.3 mM/d at pH 8.7 to 0.2 mM/d at pH 9.4,  
 500 i.e. a decrease of 86%. This was also observed for the SRR, with a decrease of 89%. The impact of pH  
 501 on POR and SRR was also observed after introduction of the cement paste and stabilization of the  
 502 operating conditions (especially pH), with a decrease in POR of 83% (2.9 at pH 9.0 and 0.5 mM/d at  
 503 pH 9.4) and a decrease of 84% for SRR (1.9 mM/d at pH 9.0 and 0.3 mM/d at pH 9.4).

504 **Table 2:** SRR and POR calculated at different pH measurements with a ratio of 92 cm<sup>2</sup>/L, for the  
 505 bioreactor before the addition of MA cement pastes, and after their addition into the bioreactor. The  
 506 initial pH value corresponds to the first pH measurement at the beginning of the experiment.

|  | Solid/liquid ratio    | Average initial pH* | POR (mM/d) | Acetate production rate (mM/d) | SRR (mM/d) |
|--|-----------------------|---------------------|------------|--------------------------------|------------|
| Without MA cement paste coupon - all reactors included (+sd) |                       | 8.7 ± 0.0           | 1.3 ± 0.8  | 0.8 ± 0.1                      | 0.9 ± 0.6  |
|  |                       | 9.3 ± 0.0           | 0.8 ± 0.1  | 0.9 ± 0.1                      | 0.5 ± 0.1  |
|  |                       | 9.4 ± 0.0           | 0.2 ± 0.1  | 0.1 ± 0.1                      | 0.1 ± 0.1  |
| With MA cement paste coupon all reactors included (+sd)**    | 92 cm <sup>2</sup> /L | 9                   | 2.9        | 2.8                            | 1.9        |
|  |                       | 9.2                 | 1.5        | 2                              | 1.1        |
|  |                       | 9.3 ± 0.0           | 1.1 ± 0.8  | 1.4 ± 0.9                      | 0.8 ± 0.6  |
|  |                       | 9.4                 | 0.5        | 0.4                            | 0.3        |

507  
 508 \* pH measured after supplementation and N<sub>2</sub> bubbling; \*\* standard deviation (sd) when the calculation is possible i.e.  
 509 when there were more than three measuring points \*\*\* SRR: sulfate reduction rate; POR: propionate oxidation rate

### 510 3.2.2 Biological investigations

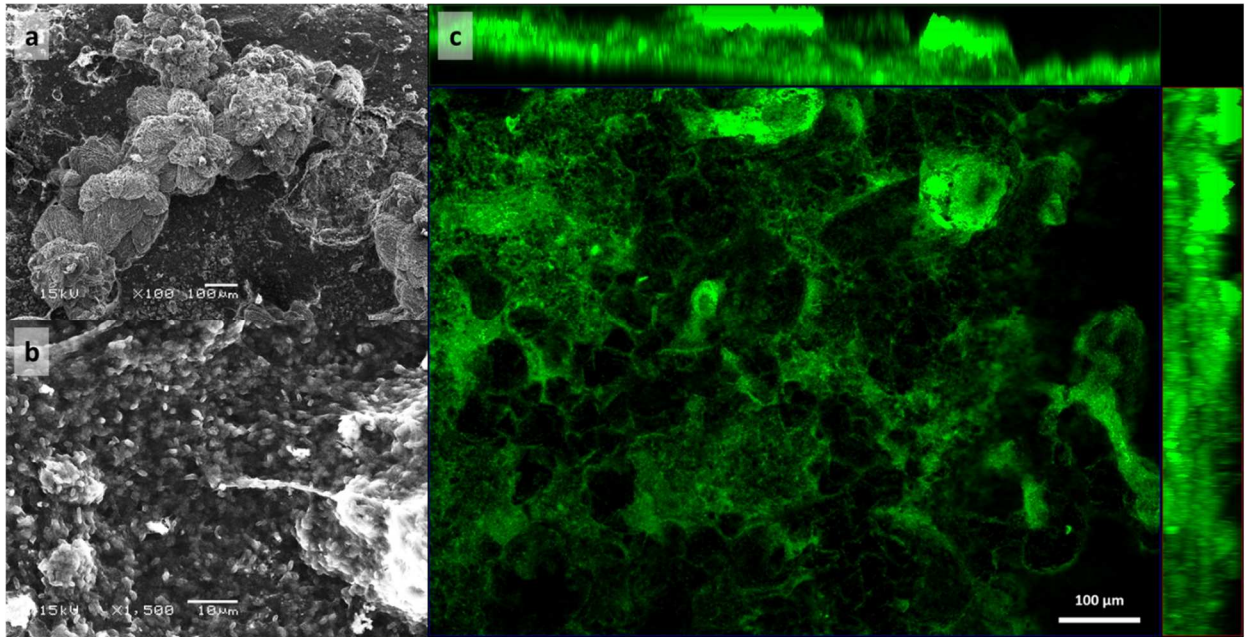
#### 511 3.2.2.1 Presence of bacteria attached to the surface

512 In order to confirm the presence of microorganisms attached to the surface and to study their  
 513 metabolic activity independently of the planktonic fraction (microorganisms in the supernatant), at

514 the end of step 3 for the experiment with MA cement paste coupons, one cement coupon of each  
515 bioreactor was transferred into a new bioreactor containing cement leachate supplemented with  
516 sulfate and propionate. This experiment was therefore conducted in duplicate (section S.1 in  
517 supplementary data). Before any biological activity, growth or redox reaction could be observed,  
518 over the two bioreactors, a transitional period before pH stabilisation of the order of one hundred  
519 days with a frequent pH adjustment (once or twice a week during the first fifty days) was necessary.  
520 This strong increase of the pH resulted from a release of the hydroxide ions still present in the  
521 cement paste (Figure S.2). After this acclimatization period, the OD increased slowly and reached the  
522 maximal value of 0.2 after 175 days. Then a significant reduction of the sulfate associated with an  
523 oxidation of the propionate into acetate and CO<sub>2</sub> was observed (Figure S.14). On average, the SRRs  
524 observed were  $0.7 \pm 0.2$  mM/d and the PORs were  $1.1 \pm 0.3$  mM/d for an average pH of 8.8.

525 A recolonization of the supernatant by microorganisms reducing the sulfate and oxidizing the  
526 propionate was established. The cement paste being the only source of microorganisms, the  
527 presence of bacteria attached to the surface was most likely.

528 Therefore, at the end of this experiment, microscopic observations were performed: epifluorescence  
529 microscopy observations on the cement surface after DNA labelling by Syto 9, and SEM observations  
530 (Figure 5). Both techniques revealed bacteria adhered to the surface of the MA cement paste coupon  
531 around the calcite precipitates (Figure 5). EDS analyses (data not shown) detected phosphorus in  
532 spherical structures. The morphology of which also observed in Figure 5.b, suggested that they were  
533 bacteria.



534

535 **Figure 5:** SEM (a,b) and epifluorescence microscopic imaging (c) with Syto 9 labelling of an MA  
 536 cement paste after 180 days of immersion in the bioreactor after renewal of the culture medium by  
 537 cement leachate enriched with sulfate and propionate.

538

### 539 3.2.2.2 Evolution of bacterial communities

540 The microbial populations from the inoculum and present in the bioreactor were identified by  
 541 molecular inventory of 16S rRNA (see section S.2 and Figure S.9 in supplementary data). The analysis  
 542 of the microbial populations from the inoculum, i.e. activated sludge, showed that the main phyla  
 543 within the microbial community of the inoculum were Proteobacteria ( $\approx 20\%$ ), Actinobacteria (10%),  
 544 Planctomycetes (9%), Bacteroidetes (5%), Chloroflexi (4%) and Firmicutes (3%) (Figure S.8).

545 Before MA cement paste coupon addition, the community was dominated by three phyla:  
 546 *Proteobacteria* (26%), *Firmicutes* (29%) and *Chlorobi* (22%). The phylum *Proteobacteria* was mostly  
 547 represented by the class *Deltaproteobacteria* (11.5%) of which the genus *Desulfobulbus* was present  
 548 at 5 % and the others were unknown. The genus *Candidatus Competibacter* made up 6% of the class  
 549 *Gammaproteobacteria* and the remaining 8% corresponded to species belonging to the orders  
 550 Burkholderiales, Rhodobacterales and Rhizobiales, having less than 1% representability. The phylum  
 551 *Firmicutes* (29%) was represented by *Fusibacter* (27%) and the phylum *Chlorobi* (22%) was  
 552 represented by *Ignavibacterium*.

553 After 71 days of culture, 45 of which in the presence of MA cement paste coupon, the community  
554 was dominated by the phylum *Proteobacteria* (83%), especially the species *Desulfomicrobium sp.* The  
555 medium was then renewed with fresh cement leachate supplemented by sulfate and propionate. The  
556 supernatant was recolonized after 180 days of culture. This population was analysed but 51% of the  
557 sequences obtained could not be characterized. The other sequences were mostly attributed to the  
558 phylum *Proteobacteria* (45%), especially the species *Desulfomicrobium sp.*

### 559 3.2.3 Analysis of the cement coupons

#### 560 3.2.3.1 Macroscopic observations

561 Compared to the advanced ageing protocol, the moderate ageing process did not visibly fracture the  
562 surfaces of the MA cement paste coupons, which maintained a homogenous light grey pattern  
563 (slightly lighter tint than that of a sound specimen) (Figure S.11.e). However, dark spots were noticed  
564 on cement paste surfaces after 45 days of immersion in the bioreactors enriched with sulfate-  
565 reducing and propionate-oxidizing bacteria (Figure S.11 f).

566

#### 567 3.2.3.2 Chemical and microstructural characterization of MA cement pastes coupons before and after 568 their exposure in the bioreactors

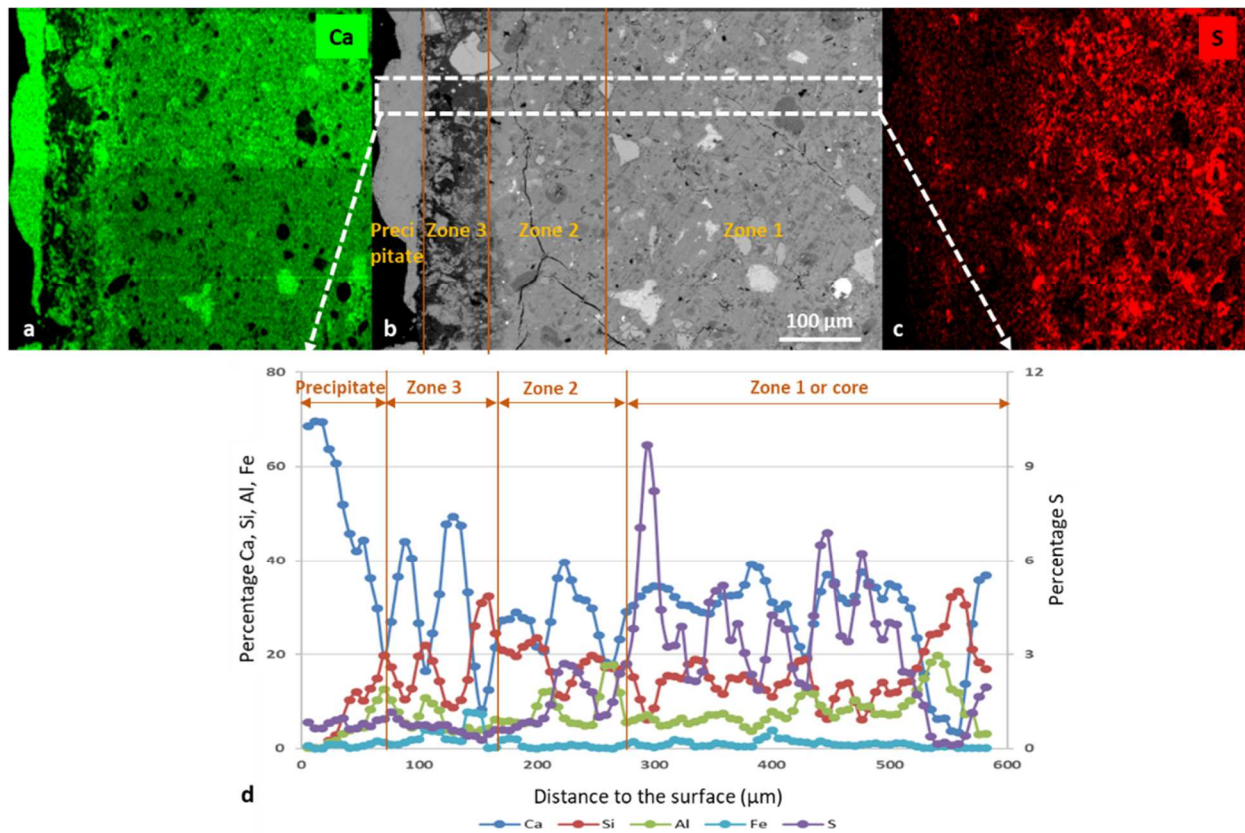
##### 569 • **MA cement paste before exposure:**

570 Figure S.15 in the supplementary data presents SEM observations (Figure S.15.a) and a chemical  
571 composition profile (Figure S.15) obtained with EDS of an MA cement paste coupon after the  
572 moderate ageing (MA) protocol with pure water. The analyses reveal a much less intense  
573 deterioration pattern induced by the moderate ageing process (Figure S.15) compared to the  
574 advanced ageing process (Figure 3). Two zones were visible: zone 1 or the core of the specimen,  
575 where the average calcium content was 34.8%, and zone 2, only slightly decalcified: the content of Ca  
576 was 28.6% in this zone. The average sulfur (S) content in zone 1 was 1.8%, but it dropped to 0.4% in  
577 zone 2. Si, Al and Fe were preserved in this zone, which had a thickness of about 150  $\mu\text{m}$ .

578

##### 579 • **MA cement paste coupon after exposure to the bioreactor:**

580 The SEM imaging of the cross section of MA cement paste coupon after 45 days of immersion in the  
581 bioreactor allowed three different zones to be distinguished (Figure 6.b).



582  
583 **Figure 6:** Scanning electron micrograph (SEM), in backscattered electron mode, of the external layer  
584 from the cross section of a MA cement paste coupon that was immersed for 45 days in the  
585 bioreactor (b), and EDS mapping of the calcium content in green (a) and sulfur content in red (c). In  
586 (d), evolution of the chemical composition (Ca, Si, Al, Fe, and S) of zones 1, 2, 3 and 4 identified  
587 according to the distance to the surface of MA cement paste coupons. Zone 1 corresponds to the  
588 core of the MA cement paste, while zone 3 corresponds to its surface that was exposed to the  
589 microbial communities.

590  
591 The microstructural, chemical and mineralogical patterns of zone 1 correspond to those of a sound  
592 hydrated cement paste. Zone 2, with a thickness of 110 μm, contained fewer anhydrous grains, but  
593 the average density of the hydrated matrix seemed equivalent to that of zone 1. Coupling these  
594 results with EDS analysis determined that the calcium contents in zones 1 and 2 were similar (27.7%  
595 and 28.36 %, respectively) (Figure 6.a). The calcium content decreased considerably at the limit  
596 between zone 2 and zone 3 and reached 10.5%, due to Ca leaching from the cement paste. Despite  
597 the ongoing process of calcium leaching from MA cement paste coupons, the calcium content in zone

3, although irregular, was high. This higher calcium content could have resulted from the microbial activity of the SRBs in the bioreactor (Figure 6.a), whose CO<sub>2</sub> production could have reacted with the Ca released from the pulp and led to the precipitation of CaCO<sub>3</sub> in this area. EDS Ca mapping (Figure 6.a), SEM observation in back-scattered electron mode (Figure 6.b), and the EDS profile (Figure 6. d) highlighted the presence of a precipitate a few tens of μm thick and mainly composed of calcium on the surface of the coupon, likely a CaCO<sub>3</sub> precipitate according data obtained by XRD (data not shown). Regarding the fate of sulfur, the average content of this element in zone 1 was 3.1%. It decreased in zone 2 and even more near the surface (zone 3) but EDS S mapping showed areas with local enrichment in sulfur, even close to the surface (Figure 6.c). The SEM imaging coupled to EDS analyses highlighted several cracks with such localized sulfur enrichment (Figure 6.b, c, and in supplementary data S.16 with the Table S.4). Their microstructure suggested the presence of secondary expansive ettringite, which could explain the sulfur enrichment. In zone 3, the sulfur content stabilized around 1.1%. The presence of sulfur in zone 3, as well as the precipitation of secondary ettringite in zone 2, could be related to a diffusion of exogenous sulfur present in the bioreactor. The total thickness of zones 2 and 3 was about 210 μm.

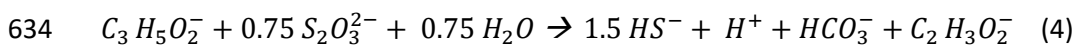
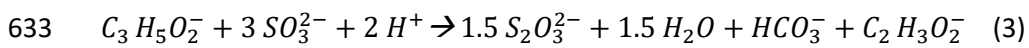
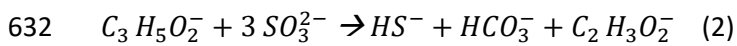
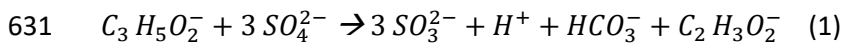
SEM imaging coupled with EDS analysis of the surface of the specimens highlighted several calcium-rich precipitates with different morphologies and sizes from ten to a hundred μm (Figure S.17 and Table S.5 in supplementary data), suggesting the presence of calcium carbonate precipitates (CaCO<sub>3</sub>). Other punctual chemical analyses performed on the cement paste surface (Table S.5) revealed compounds mainly composed of aluminium and silicon, which probably correspond to silico-aluminous fly ash contained in CEM V cement.

## 4 Discussion

### 4.1 Bio-reactivity of sulfate and propionate

For a pH close to 9.0, with or without cement paste, the propionate was oxidized into CO<sub>2</sub> and acetate, which accumulated in the bioreactor. The oxidation of the propionate was incomplete; i.e., propionate was not completely oxidised to CO<sub>2</sub>, an average of 74% of the oxidized carbon being

624 transformed into acetate. Moreover, the presence of sulfite and thiosulfate, two intermediate forms  
 625 of sulfate reduction, was observed. According to the sulfate reduction pathway, coupled with the  
 626 oxidation of propionate, sulfate was first reduced to sulfite (reaction 1 below) which was reduced to  
 627 sulfide (reaction 2) or to thiosulfate, which is an intermediate step in sulfite reduction (reactions 3  
 628 and 4) (Bradley et al., 2011; Qian et al., 2019). The global reaction of the sulfate reduction to sulfide  
 629 coupled to the oxidation of propionate is described by reaction 5 at pH close to 9.0, with HS<sup>-</sup> as  
 630 major form of hydrogen sulfide (Widdel and Pfennig, 1981):



636

637 According to the redox reactions, the reduction of sulfate to sulfite (reaction 1) or thiosulfate to  
 638 sulfide (reaction 4) leads to a production of proton (H<sup>+</sup>) which can lead to a decrease in pH. This was  
 639 indeed observed during the reduction of sulfate in the different reactors with and without cement  
 640 paste. On the other hand, the reduction of sulfite to thiosulfate requires a consumption of protons  
 641 (reaction 4), which, in an alkaline environment, can be a limiting step of the sulfate reduction due to  
 642 the low (bio) availability of H<sup>+</sup>.

643 Before the addition of AA or MA cement paste coupons, the PORs were between 1.3 and 2.1 mM/d  
 644 at pH close to 9.0 and the SRRs were between 0.9 and 1.4 mM/d. However, the increase of the pH  
 645 induced a decrease in the PORs and SRRs similar to an inhibiting effect, especially for pH values  
 646 above 9.3: the POR decreased by 89 % and the SRR by 85% when the pH increased from 8.7 to 9.3  
 647 without cement paste. With MA cement paste coupons (Table 2), both the SRRs and the PORs  
 648 decreased by 20% for each pH increase of 0.1, between 9.0 and 9.3, which represented an overall  
 649 decrease of 62% for POR and 68% for SRR for a pH increase from 9.0 to 9.3. For pH higher than 9.3,

650 the SRRs and PORs decreased, respectively, by 83% and 84% compared to the values obtained at pH  
651 9.0 and no significant sulfate reduction was observed for pH above 9.5. Other studies have also  
652 noted a poor reduction of sulfate at pH values above 10 and have defined a critical pH value of 9.3  
653 above which sulfate reduction is strongly affected (Goeres et al., 1998; Rizoulis et al., 2012). In the  
654 case of AA cement paste coupons addition, the pH impact on the PORs and SRRs was also observed  
655 but was more moderate. The rates decreased by 43% for POR and 39% for SRR for a pH increase from  
656 9.0 to 9.4. This can be explained by the higher level of alteration of AA cement paste coupons and  
657 the presence of lower SA/V ratio, and therefore a lower alkaline cation and hydroxide reserve.

658 The propionate oxidation and sulfate reduction process together with the associated ratios, PORs  
659 and SRRs, were little impacted by the addition of cement paste once the pH was stabilized and the  
660 microbial activity in place for SA/V ratio below 96 cm<sup>2</sup>/L. An increase in residual thiosulfate was  
661 observed in the liquid phase, with an average concentration of 4 mM, i.e. twice as high as before the  
662 cement paste addition. However, the addition of AA type cement paste coupons with a SA/V ratio of  
663 96 cm<sup>2</sup>/L had an impact on SRRs and PORs by inducing a decrease of about 35% compared to the  
664 performance obtained without cement paste. The PORs and SRRs obtained at pH 9.2 with a SA/V  
665 ratio of 92 cm<sup>2</sup>/L were similar between experiments with AA and MA cement paste coupons.  
666 Moderate ageing of the cement paste delayed the emergence of microbial activity by inducing a  
667 strong increase in the pH of the medium, but once the pH stabilized and the microbial activity  
668 established, the ageing of the cement paste did not strongly affect PORs or SRRs at pH 9.2.

669 Concerning models of the geochemical evolution of storage, these data will allow us to define the  
670 conditions in which an oxidation of the organic matter coupled with the reduction of the sulfate  
671 could be included in the geochemical equations used for the safety evaluation (Bagaria et al., 2021) ,  
672 for pH lower than 10 or 9.5 and after alteration of the cement matrix. Moreover, the ability to specify  
673 a pH threshold above which no microbial activity, thus no sulfate reduction nor organic matter  
674 oxidation, occurs is essential. In sum, propionate oxidation and sulfate reduction was strongly slowed  
675 down for pH higher than 9.3 and was not observed for pH above 9.5.

676

#### 677 **4.2 Cement paste pre-treatment**

678 The AA cement paste coupons process induced strong alteration that was expressed by cracking of  
679 the outer layer and a marked chemical and microstructural zonation. The deterioration patterns  
680 were typical of the alteration of cement pastes exposed to acetic acid (Bertron et al., 2005b;  
681 Duchesne and Bertron, 2013) with a fast leaching of calcium ( $\text{Ca}^{2+}$ ), hydroxide ( $\text{OH}^-$ ) and alkaline ( $\text{Na}^+$ ,  
682  $\text{K}^+$ ) ions without promotion of any secondary precipitation or any specific interactions between the  
683 acetate anion ( $\text{CH}_3\text{COO}^-$ ) and the cations leached from cement pastes (Bertron et al., 2007, 2005a,  
684 2005b; Bertron and Duchesne, 2013). The outermost zone was strongly decalcified. This intense  
685 leaching of calcium caused chemical shrinkage that resulted in the dense cracking pattern of the  
686 outer layer, visible on the cement paste surface (Bertron et al., 2007).

687 The moderate ageing (MA) process, which consisted of one week leaching of the cement paste with  
688 ultrapure water, induced much less intense alteration of the cementitious specimens, and thus  
689 higher pH after their introduction into the microbial medium. The specimens maintained a  
690 homogeneous light grey pattern close to that of a sound specimen, without any visible cracks. The  
691 depth of alteration (150  $\mu\text{m}$ ) was almost ten times less than that of the AA cement paste coupons  
692 and the chemical composition of the cement paste in contact with the liquid medium was little  
693 affected by the leaching (slight decalcification and sulfur leaching).

#### 694 **4.3 Impact of biological activity on solid matrices**

695 The exposure of the cement pastes to microbial activity, which catalysed the reduction of sulfate to  
696 sulfide coupled with an oxidation of propionate to  $\text{HCO}_3^-$  and acetate, induced strong interaction  
697 between microorganisms and cementitious material. These interactions led to an increase in the  
698 degradation front depth of the outer layer with respect to the initial states of the AA and MA cement  
699 paste coupons after their pre-ageing treatment, particularly for the AA pastes, which were initially  
700 the most altered. For the AA cement paste coupons, the depth of degradation to the outer layer was  
701 670  $\mu\text{m}$  after pre-treatment and increased to 750  $\mu\text{m}$  after exposure to the biological activity of

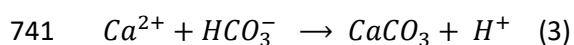
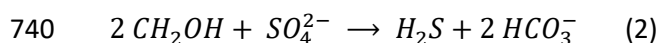
702 propionate oxidation and sulfate reduction whose reaction induced a production of sulfide and  
703 carbon dioxide. The specimen outer layer was progressively decalcified and the calcium content  
704 dropped to zero at the surface of the AA cement paste (vs. 8% Ca at the surface of AA paste after the  
705 pre-ageing process). The intensified decalcification of the AA cement paste coupons in the already  
706 altered zones may have been the consequence of aggressive chemical species produced by the SRB,  
707 notably the protons or acetate associated with the sulfate reduction and propionate oxidation  
708 (reactions 5), which induced an additional acid attack on the matrix (Magniont et al., 2011; Voegel et  
709 al., 2019b, 2016).

710 The presence of black spots or black stains was observed on AA cement pastes coupons surface after  
711 18 days of immersion, and after 45 days on MA cement paste surface. These black spots might have  
712 been due to the precipitation of iron sulfide minerals resulting from the reactions between Fe(II) of  
713 the cement paste and sulfide formed by the microbial reduction of sulfate in solution (Amen et al.,  
714 2018; Etim et al., 2021). This reaction seems to have been more marked in the case of AA cement  
715 pastes coupons because of their higher level of alteration, which may have facilitated access to the  
716 ferrous sites.

717 In the transition zone between the outer layer exposed to the medium and the sound core of the  
718 cement paste, the presence of several cracks containing S, Ca and Al was observed for the MA  
719 cement pastes coupons. This is typical of secondary expansive ettringite precipitation, which may  
720 have occurred by reaction of exogenous non-reduced sulfate, diffused from the microbial medium  
721 through the outer zone, with calcium aluminate phases in the cement matrix (Menéndez et al.,  
722 2013).

723 Noticeable secondary precipitation of Ca-rich minerals was also observed in the outer layer and/or  
724 on the surface of AA and MA cement paste coupons exposed to SRB activity. A crack network sealed  
725 by calcium-rich secondary precipitates (mostly  $\text{CaCO}_3$ ) was observed in the altered zone of AA  
726 cement paste coupons. Moreover, Ca-rich precipitates with different morphologies and sizes (from  
727 ten to a hundred  $\mu\text{m}$ ), typical of calcium carbonates, were also observed on the surface of the AA

728 and MA cement paste coupons. The precipitation of  $\text{CaCO}_3$  in the cracks and on the surface can be  
729 viewed as self-healing by microbial induced calcium carbonate precipitation, which has been widely  
730 studied in other contexts (Ersan et al., 2018; Joshi et al., 2017; Vijay et al., 2017). It occurs from the  
731 reaction of the Ca leached from the cement matrix with the microbially produced  $\text{CO}_2$ . Actually,  
732 bacterial walls, being negatively charged, have a strong affinity for nearby cations, including calcium  
733 ( $\text{Ca}^{2+}$ ), which tend to bind to their cell walls (Vijay et al., 2017).  $\text{Ca}^{2+}$  ions react with  $\text{CO}_2$  produced by  
734 the microbial oxidation of organic matter during sulfate reduction and then generate calcite  
735 precipitation around the bacterial cell surface, which serves as a nucleation site (Ersan et al., 2018;  
736 Joshi et al., 2017). The formation of calcium carbonate precipitate, in the polymorphic form of calcite  
737 or aragonite, by sulfate-reducing bacteria has been observed previously (Hammes and Verstraete,  
738 2002; Joshi et al., 2017) and the following reactions (2), with methanol as electron donor, and (3)  
739 have been described by Peckmann et al. (1999):



742 In the case of propionate oxidation to acetate, only one molecule of bicarbonate was produced  
743 (Reaction 5). Thus, for each molecule of propionate oxidized, one molecule of calcium was consumed  
744 and, theoretically, one molecule of calcite was produced.

745 Microbial colonization sites on solid materials are usually preferentially located in areas with good  
746 bio-receptivity, where surfaces are less chemically and pH aggressive and are rough enough to  
747 provide protection from an external environment that may be chemically or mechanically aggressive,  
748 while ensuring good access to the substrate, (Flemming and Wingender, 2010; Voegel et al., 2016).  
749 Based on these criteria, the initial bacterial adhesion, at the origin of the biofilm formation, could  
750 preferentially take place on weathered areas or in cracks. This colonization and localized high  
751 concentrations of SRB can lead to accelerated deterioration (such as calcification) due to the  
752 production of aggressive metabolites. However, it can also lead to a consolidation of these areas by  
753 precipitation of calcium, in the form of calcite, within the cracks or by forming a protective layer of

754 calcite on the surface. This would limit the subsequent diffusion of residual calcium from the paste to  
755 the outside.

756 In the context of geological storage, a catalysis of sulfate reduction coupled with an oxidation of the  
757 organic matter is assumed. These reactions could lead to a production of sulfides, organic acids (e.g.  
758 acetate) and/or CO<sub>2</sub>, thus acidifying the system and inducing an attack of the cement materials.

759

#### 760 **4.4 Cement paste colonisation by microorganisms**

761 The analysis of the microbial population before the introduction of MA cement pastes coupons has  
762 indeed enabled the identification of bacterial species belonging to the genus *Desulfobulbus*,  
763 *Competibacter*, *Ignavibacterium* and *Fusibacter*. The species belonging to the genus *Desulfobulbus*  
764 *sp.* are known to be SRB that use propionate as an electron donor and a carbon source to produce  
765 and reduce thiosulfate (El Houari et al., 2017; Sorokin et al., 2014; Widdel and Pfennig, 1981). Some  
766 species, such as *Desulfobulbus alkaliphilus sp.*, are obligately alkaliphilic and grow at pH 8.2 – 10.2  
767 with an optimum at pH 9.4 (Sorokin et al., 2012). Concerning the species from the genus *Fusibacter*,  
768 *Competibacter* or *Ignavibacterium*, little information about their role in the sulfate reduction process  
769 is available in the literature. Some species from the genus *Ignavibacterium* have been reported as  
770 versatile and able to reduce sulfate, e.g. *Ignavibacterium album* (Lino et al., 2010). No *Fusibacter* has  
771 been reported to be involved in sulfate reduction but some species may participate in sulfite or  
772 thiosulfate reduction and can thus be considered as alkaliphilic bacteria; i.e. bacteria that can live  
773 and thrive in an alkaline environment with pH ranging from 8.5 to 11. (Liu et al., 2021; Ravot et al.,  
774 2015).

775 In addition to the microbial population present in the liquid phase, the population of microorganisms  
776 adhered to the surface of the cement paste has also been investigated. The presence of  
777 microorganisms attached to the surface of the cement paste has been confirmed by microscopic  
778 observation (epifluorescence and SEM) and experimentally (transfer of the cement paste in contact  
779 with the inoculum into a fresh medium without microorganism (Durban et al., 2020)). The coverage

780 rate or the surface area colonized by microorganisms according to the different levels of cement  
781 paste ageing (MA or AA cement paste coupons) could not be studied. However, the time required to  
782 observe biological activity after transferring the cement pastes to a new reactor: 55 days for MA  
783 cement paste coupons versus 1 day for AA, suggests that the AA cement paste coupons were more  
784 easily and/or densely colonized than the MA cement pastes. The latter, less altered, released more  
785 alkaline ion and hydroxide, imposing a pH that was not favourable to the growth and biological  
786 activity of SRB. Whatever the degree of alteration and the culture time, the same species was  
787 identified at the end of the experiments: *Desulfomicrobium sp.*. *Desulfomicrobium sp.* is a SRB that  
788 grows in anaerobic environments and uses sulfate or sulfoxyanions as terminal electron acceptors,  
789 producing H<sub>2</sub>S, coupled with the oxidation of simple organic compounds such as lactate or  
790 propionate, whose oxidation is, in some cases, incomplete and produces acetate as well as CO<sub>2</sub>  
791 (Genthner and Devereux, 2015). This SRB strain catalyses sulfate reduction under pH close to 8.5, i.e.  
792 less sensitive to moderately alkaline pH than other SRB, such as *Desulfovibrio sp.* – a frequently  
793 occurring SRB better adapted to reducing sulfate under acidic or neutral environments (Zheng et al.,  
794 2014). Several works by Sorokin et al. (2015, 2014) have shown that some SRBs from soda lakes, such  
795 as *Desulfonatronum sp.*, *Desulfonatronovibrio sp.* or *Desulfobulbus sp.*, could, in the presence of  
796 propionate, reduce sulfate at pH close to 11. However, these species were not selected in the  
797 context of our study, although *Desulfobulbus sp.* was identified in the liquid phase before the  
798 introduction of the MA cement paste. The addition of AA and MA cement paste coupons into the  
799 bioreactor provided a niche environment for the SRB community to establish colonies or biofilm  
800 within the altered surfaces.

801 Increasing the SA/V ratio increases the surface area available to microorganisms to develop and  
802 would therefore promote microbial activity (growth and redox reaction) and the alteration of  
803 cementitious materials. Furthermore, due to the leaching of cementitious materials, the increase in  
804 the SA/V ratio will also favour an alkalisation of the environment with more alkaline pH values,  
805 which will have a limiting impact on biological activity. There are therefore several competing

806 phenomena. This study has shown that colonisation of the material is slow but seems to be favoured  
807 when the cementitious materials are altered. After transferring the cementitious material into a new  
808 reactor, on the one hand a recolonisation of the supernatant was observed and on the other hand it  
809 took at least 15 days of cultivation to reach SRR and POR comparable to the initial reactor. Thus,  
810 there were active microorganisms on the cement matrix.

## 811 **5. Conclusion**

812 This study highlights the possible organic matters oxidation, such as propionate coupled with the  
813 sulfate reduction by microbial activities for cementitious systems comparable to those found in  
814 nuclear waste cells. The use of different ratios of solid cement paste surface area to liquid volume  
815 generated different pH value and in different cementitious environments to be investigated. To  
816 mimic short-term and long-term cement ageing in the nuclear waste repository, moderate ageing  
817 (MA) and advanced ageing (AA) techniques were applied. The study allowed to determine the sulfate  
818 reduction and propionate oxidation rates at different pH values – around 1.6 and 1.4 mM  $\text{SO}_4^{2-}$ /d and  
819 1.8 and 2.1 mM propionate/d at pH close to 9.0, with or without cement paste, respectively. It also  
820 highlights the strong decrease of the sulfate reduction/propionate oxidation rate with increasing pH:  
821 i.e. for a pH increase from 9.0 to 9.4 a decrease of 22% and 84% with AA and MA cement pastes  
822 coupons, respectively, and for a pH increase from 8.7 to 9.4 a decrease of 89% without cement paste.  
823 No sulfate or propionate oxidation was observed for pH above 9.5. The presence of microorganisms  
824 attached to the surface of cement pastes has been confirmed. This presence induced strong  
825 biogeochemical interactions between microorganism and cementitious material : microbial activity  
826 near the surfaces of solid materials led not only to alterations such as decalcification and sulfide  
827 enrichment with the appearance of secondary ettringite, particularly on MA pastes, but also to a  
828 relocation of calcium in the form of calcite either within the micro-fracture (AA cement paste  
829 coupons) or on the surface of the material (MA cements paste coupons) comparable to self-healing  
830 phenomena.

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## 835 **Credit authorship statement**

836 Nadège Durban: Conceptualisation, Methodology, Investigation, Validation, Formal analysis, Writing -  
837 Original Draft, Writing - Review & Editing, Visualization.

838 Alexandra Bertron: Conceptualisation, Methodology, Ressources, Writing - Original Draft, Writing -  
839 Review & Editing, Supervision, Project administration, Funding acquisition.

840 Vanessa Sonois-Mazars: Validation, Investigation.

841 Maud Schiettekatte: Validation, Investigation.

842 Gerald Matar: Writing - Review & Editing.

843 Pierre Albina: Methodology, Validation.

844 Achim Albrecht: Conceptualisation, Supervision, Writing - Review & Editing

845 Jean-Charles Robinet: Conceptualisation, Project administration

846 Benjamin Erable: Conceptualisation, Methodology, Ressources, Supervision, Writing - Review &  
847 Editing Project administration

848

## 849 **References**

850 Abrahamsen, L., Arnold, T., Brinkmann, H., Leys, N., Merroun, M., Mijndonckx, K., Moll, H., Polvika,  
851 P., Ševců, A., Small, J., Vikman, M., Wouters, K., 2015. A Review of Anthropogenic organic  
852 Wastes and Their Degradation Behaviour. Microbiol. Nucl. Waste Dispos. (MIND deliverable  
853 D1.1.).

854 Albina, P., Durban, N., Bertron, A., Albrecht, A., Robinet, J.-C., Erable, B., 2021. Nitrate and nitrite  
855 bacterial reduction at alkaline pH and high nitrate concentrations, comparison of acetate

856 versus dihydrogen as electron donors. *J. Environ. Manage.* 280, 111859.  
857 <https://doi.org/10.1016/j.jenvman.2020.111859>

858 Albrecht, A., Bertron, A., Libert, M., 2013. Microbial Catalysis of Redox Reactions in Concrete Cells of  
859 Nuclear Waste Repositories: A Review and Introduction, in: Bart, F., Cau-di-Coumes, C.,  
860 Frizon, F., Lorente, S. (Eds.), *Cement-Based Materials for Nuclear Waste Storage*. Springer  
861 New York, New York, NY, pp. 147–159. [https://doi.org/10.1007/978-1-4614-3445-0\\_14](https://doi.org/10.1007/978-1-4614-3445-0_14)

862 Alquier, M., Kassim, C., Bertron, A., Sablayrolles, C., Rafrafi, Y., Albrecht, A., Erable, B., 2014.  
863 *Halomonas desiderata* as a bacterial model to predict the possible biological nitrate  
864 reduction in concrete cells of nuclear waste disposals. *J. Environ. Manage.* 132, 32–41.  
865 <https://doi.org/10.1016/j.jenvman.2013.10.013>

866 Amen, T.W.M., Eljamal, O., Khalil, A.M.E., Matsunaga, N., 2018. Evaluation of sulfate-containing  
867 sludge stabilization and the alleviation of methanogenesis inhibition at mesophilic  
868 temperature. *J. Water Process Eng.* 25, 212–221. <https://doi.org/10.1016/j.jwpe.2018.08.004>

869 Bagaria, F., Riba, O., Albrecht, A., Robinet, J.-C., Madé, B., Román-Ross, G., 2021. Predicting  
870 degradation of organic molecules in cementitious media. *Prog. Nucl. Energy* 140, 103888.  
871 <https://doi.org/10.1016/j.pnucene.2021.103888>

872 Bary, B., 2008. Simplified coupled chemo-mechanical modeling of cement pastes behavior subjected  
873 to combined leaching and external sulfate attack. *Int. J. Numer. Anal. Methods Geomech.* 32,  
874 1791–1816. <https://doi.org/10.1002/nag.696>

875 Bertron, A., 2014. Understanding interactions between cementitious materials and microorganisms:  
876 a key to sustainable and safe concrete structures in various contexts. *Mater. Struct.* 47,  
877 1787–1806. <https://doi.org/10.1617/s11527-014-0433-1>

878 Bertron, A., Duchesne, J., 2013. Attack of Cementitious Materials by Organic Acids in Agricultural and  
879 Agrofood Effluents, in: Alexander, M., Bertron, A., De Belie, N. (Eds.), *Performance of*  
880 *Cement-Based Materials in Aggressive Aqueous Environments*. Springer Netherlands,  
881 Dordrecht, pp. 131–173. [https://doi.org/10.1007/978-94-007-5413-3\\_6](https://doi.org/10.1007/978-94-007-5413-3_6)

882 Bertron, A., Duchesne, J., Escadeillas, G., 2007. Degradation of cement pastes by organic acids.  
883 *Mater. Struct.* 40, 341–354. <https://doi.org/10.1617/s11527-006-9110-3>

884 Bertron, A., Duchesne, J., Escadeillas, G., 2005a. Accelerated tests of hardened cement pastes  
885 alteration by organic acids: analysis of the pH effect. *Cem. Concr. Res.* 35, 155–166.  
886 <https://doi.org/10.1016/j.cemconres.2004.09.009>

887 Bertron, A., Duchesne, J., Escadeillas, G., 2005b. Attack of cement pastes exposed to organic acids in  
888 manure. *Cem. Concr. Compos.* 27, 898–909.  
889 <https://doi.org/10.1016/j.cemconcomp.2005.06.003>

890 Bertron, A., Escadeillas, G., de Parseval, P., Duchesne, J., 2009. Processing of electron microprobe  
891 data from the analysis of altered cementitious materials. *Cem. Concr. Res.* 39, 929–935.  
892 <https://doi.org/10.1016/j.cemconres.2009.06.011>

893 Bradley, A.S., Leavitt, W.D., Johnston, D.T., 2011. Revisiting the dissimilatory sulfate reduction  
894 pathway. *Geobiology* 9, 446–457. <https://doi.org/10.1111/j.1472-4669.2011.00292.x>

895 Chang, Y.C., Le Puil, M., Biggerstaff, J., Randall, A.A., Schulte, A., Taylor, J.S., 2003. Direct estimation  
896 of biofilm density on different pipe material coupons using a specific DNA-probe. *Mol. Cell.*  
897 *Probes* 17, 237–243. <https://doi.org/10.1016/j.mcp.2003.07.004>

898 Chen, Y., Ruhyadi, R., Huang, J., Yan, W., Wang, G., Shen, N., Hanggoro, W., 2021. Comprehensive  
899 comparison of acidic and alkaline anaerobic fermentations of waste activated sludge.  
900 *Bioresour. Technol.* 323, 124613. <https://doi.org/10.1016/j.biortech.2020.124613>

901 Dazères, A., Le Bescop, P., Cau-Dit-Coumes, C., Brunet, F., Bourbon, X., Timonen, J., Voutilainen, M.,  
902 Chomat, L., Sardini, P., 2014. On the physico-chemical evolution of low-pH and CEM I cement  
903 pastes interacting with Callovo-Oxfordian pore water under its in situ CO<sub>2</sub> partial pressure.  
904 *Cem. Concr. Res.* 58, 76–88. <https://doi.org/10.1016/j.cemconres.2014.01.010>

905 Davis, J.A., 1984. Complexation of trace metals by adsorbed natural organic matter. *Geochim.*  
906 *Cosmochim. Acta* 48, 679–691. [https://doi.org/10.1016/0016-7037\(84\)90095-4](https://doi.org/10.1016/0016-7037(84)90095-4)

907 Duchesne, J., Bertron, A., 2013. Leaching of Cementitious Materials by Pure Water and Strong Acids  
908 (HCl and HNO<sub>3</sub>), in: Alexander, M., Bertron, A., De Belie, N. (Eds.), Performance of Cement-  
909 Based Materials in Aggressive Aqueous Environments. Springer Netherlands, Dordrecht, pp.  
910 91–112. [https://doi.org/10.1007/978-94-007-5413-3\\_4](https://doi.org/10.1007/978-94-007-5413-3_4)

911 Duran, N., Seabra, A., 2012. Microbial Syntheses of Metallic Sulfide Nanoparticles: An Overview. *Curr.*  
912 *Biotechnol. E* 1, 287–296. <https://doi.org/10.2174/2211550111201040287>

913 Durban, N., Rafrafi, Y., Rizoulis, A., Albrecht, A., Robinet, J.-C., Lloyd, J.R., Bertron, A., Erable, B., 2018.  
914 Nitrate and nitrite reduction at high pH in a cementitious environment by a microbial  
915 microcosm. *Int. Biodeterior. Biodegrad.* 134, 93–102.  
916 <https://doi.org/10.1016/j.ibiod.2018.08.009>

917 Durban, N., Sonois-Mazars, V., Albina, P., Bertron, A., Albrecht, A., Robinet, J.-C., Erable, B., 2020.  
918 Nitrate and nitrite reduction activity of activated sludge microcosm in a highly alkaline  
919 environment with solid cementitious material. *Int. Biodeterior. Biodegrad.* 151, 104971.  
920 <https://doi.org/10.1016/j.ibiod.2020.104971>

921 El Houari, A., Ranchou-Peyruse, M., Ranchou-Peyruse, A., Dakdaki, A., Guignard, M., Idouhammou, L.,  
922 Bennisse, R., Bouterfass, R., Guyoneaud, R., Qatibi, A.-I., 2017. *Desulfobulbus oligotrophicus*  
923 sp. nov., a sulfate-reducing and propionate-oxidizing bacterium isolated from a municipal  
924 anaerobic sewage sludge digester. *Int. J. Syst. Evol. Microbiol.* 67, 275–281.  
925 <https://doi.org/10.1099/ijsem.0.001615>

926 Ersan, Y.C., Boon, N., De Belie, N., 2018. Granules with activated compact denitrifying core (ACDC) for  
927 self-healing concrete with corrosion protection functionality. Presented at the Final  
928 conference of RILEM TC 253-MCI: Microorganisms-cementitious materials interactions,  
929 RILEM Publications, pp. 475–484.

930 Etim, I.-I.N., Dong, J., Wei, J., Nan, C., Pokharel, D.B., Umoh, A.J., Xu, D., Su, M., Ke, W., 2021. Effect of  
931 organic silicon quaternary ammonium salts on mitigating corrosion of reinforced steel  
932 induced by SRB in mild alkaline simulated concrete pore solution. *J. Mater. Sci. Technol., SI:*  
933 *Advanced Corrosion-Resistant Materials and Emerging Applications* 64, 126–140.  
934 <https://doi.org/10.1016/j.jmst.2019.10.006>

935 Ewing, R.C., Whittleston, R.A., Yardley, B.W.D., 2016. Geological Disposal of Nuclear Waste: a Primer.  
936 *Elements* 12, 233–237. <https://doi.org/10.2113/gselements.12.4.233>

937 Flemming, H.C., Wingender, J., 2010. The biofilm matrix. *Nat. Rev. Microbiol.* 8, 623–633.  
938 <https://doi.org/10.1038/nrmicro2415>

939 Gallé, C., Peycelon, H., Le Bescop, P., Bejaoui, S., L’Hostis, V., Bary, B., Bouniol, P., Richet, C., 2006.  
940 Concrete long-term behaviour in the context of nuclear waste management: Experimental  
941 and modelling research strategy. *J. Phys. IV Proc.* 136, 25–38.  
942 <https://doi.org/10.1051/jp4:2006136004>

943 Genthner, B.R.S., Devereux, R., 2015. *Desulfomicrobium*, in: *Bergey’s Manual of Systematics of*  
944 *Archaea and Bacteria.* American Cancer Society, pp. 1–9.  
945 <https://doi.org/10.1002/9781118960608.gbm01032>

946 Giroudon, M., Peyre Lavigne, M., Patapy, C., Bertron, A., 2021. Blast-furnace slag cement and  
947 metakaolin based geopolymer as construction materials for liquid anaerobic digestion  
948 structures: Interactions and biodeterioration mechanisms. *Sci. Total Environ.* 750, 141518.  
949 <https://doi.org/10.1016/j.scitotenv.2020.141518>

950 Goeres, D.M., Nielsen, P.H., Smidt, H.D., Frølund, B., 1998. The effect of alkaline pH conditions on a  
951 sulphate reducing consortium from a Danish district heating plant. *Biofouling* 12, 273–286.  
952 <https://doi.org/10.1080/08927019809378360>

953 Gutierrez, O., Park, D., Sharma, K.R., Yuan, Z., 2009. Effects of long-term pH elevation on the sulfate-  
954 reducing and methanogenic activities of anaerobic sewer biofilms. *Water Res.* 43, 2549–  
955 2557. <https://doi.org/10.1016/j.watres.2009.03.008>

956 Hai, T., Lange, D., Rabus, R., Steinbüchel, A., 2004. Polyhydroxyalkanoate (PHA) Accumulation in  
957 Sulfate-Reducing Bacteria and Identification of a Class III PHA Synthase (PhaEC) in

958 Desulfococcus multivorans. Appl. Environ. Microbiol. 70, 4440–4448.  
 959 <https://doi.org/10.1128/AEM.70.8.4440-4448.2004>  
 960 Hammes, F., Verstraete, W., 2002. Key roles of pH and calcium metabolism in microbial carbonate  
 961 precipitation. Rev. Environ. Sci. Biotechnol. 1, 3–7.  
 962 <https://doi.org/10.1023/A:1015135629155>  
 963 Iino, T., Mori, K., Uchino, Y., Nakagawa, T., Harayama, S., Suzuki, K.-I., 2010. Ignavibacterium album  
 964 gen. nov., sp. nov., a moderately thermophilic anaerobic bacterium isolated from microbial  
 965 mats at a terrestrial hot spring and proposal of Ignavibacteria classis nov., for a novel lineage  
 966 at the periphery of green sulfur bacteria. Int. J. Syst. Evol. Microbiol. 60, 1376–1382.  
 967 <https://doi.org/10.1099/ijs.0.012484-0>  
 968 Joshi, S., Goyal, S., Mukherjee, A., Reddy, M.S., 2017. Microbial healing of cracks in concrete: a  
 969 review. J. Ind. Microbiol. Biotechnol. 44, 1511–1525. <https://doi.org/10.1007/s10295-017-1978-0>  
 970  
 971 Kuippers, G., Bassil, N.M., Boothman, C., Bryan, N., Lloyd, J.R., 2015. Microbial degradation of  
 972 isosaccharinic acid under conditions representative for the far field of radioactive waste  
 973 disposal facilities. Mineral. Mag. 79, 1443–1454.  
 974 <https://doi.org/10.1180/minmag.2015.079.6.19>  
 975 Lagerblad, B., Traegaardh, J., 1994. Conceptual model for concrete long time degradation in a deep  
 976 nuclear waste repository (Technical Report No. SKB-TR-95-21). Swedish Nuclear Fuel and  
 977 Waste Management Co., Stockholm (Sweden).  
 978 Larreur-Cayol, S., Bertron, A., Escadeillas, G., 2011. Degradation of cement-based materials by  
 979 various organic acids in agro-industrial waste-waters. Cem. Concr. Res. 41, 882–892.  
 980 <https://doi.org/10.1016/j.cemconres.2011.04.007>  
 981 Liu, L., Ji, J., Guo, Y., Chen, J., 2021. Use of ecological concrete for nutrient removal in coastal  
 982 sediment and its effects on sediment microbial communities. Mar. Pollut. Bull. 162, 111911.  
 983 <https://doi.org/10.1016/j.marpolbul.2020.111911>  
 984 Liu, Y., Li, X., Kang, X., Yuan, Y., Du, M., 2014. Short chain fatty acids accumulation and microbial  
 985 community succession during ultrasonic-pretreated sludge anaerobic fermentation process:  
 986 Effect of alkaline adjustment. Int. Biodeterior. Biodegrad. 94, 128–133.  
 987 <https://doi.org/10.1016/j.ibiod.2014.07.004>  
 988 Machel, H.G., 2001. Bacterial and thermochemical sulfate reduction in diagenetic settings — old and  
 989 new insights. Sediment. Geol. 140, 143–175. [https://doi.org/10.1016/S0037-0738\(00\)00176-7](https://doi.org/10.1016/S0037-0738(00)00176-7)  
 990  
 991 Magniont, C., Coutand, M., Bertron, A., Cameleyre, X., Lafforgue, C., Beaufort, S., Escadeillas, G.,  
 992 2011. A new test method to assess the bacterial deterioration of cementitious materials.  
 993 Cem. Concr. Res. 41, 429–438. <https://doi.org/10.1016/j.cemconres.2011.01.014>  
 994 Menéndez, E., Matschei, T., Glasser, F.P., 2013. Sulfate Attack of Concrete, in: Alexander, M.,  
 995 Bertron, A., De Belie, N. (Eds.), Performance of Cement-Based Materials in Aggressive  
 996 Aqueous Environments: State-of-the-Art Report, RILEM TC 211 - PAE, RILEM State-of-the-Art  
 997 Reports. Springer Netherlands, Dordrecht, pp. 7–74. [https://doi.org/10.1007/978-94-007-5413-3\\_2](https://doi.org/10.1007/978-94-007-5413-3_2)  
 998  
 999 Mu, T., Xing, J., Yang, M., 2019. Sulfate reduction by a haloalkaliphilic bench-scale sulfate-reducing  
 1000 bioreactor and its bacterial communities at different depths. Biochem. Eng. J. 147, 100–109.  
 1001 <https://doi.org/10.1016/j.bej.2019.04.008>  
 1002 Muyzer, G., Stams, A.J.M., 2008. The ecology and biotechnology of sulphate-reducing bacteria. Nat.  
 1003 Rev. Microbiol. <https://doi.org/10.1038/nrmicro1892>  
 1004 Pallud, C., Van Cappellen, P., 2006. Kinetics of microbial sulfate reduction in estuarine sediments.  
 1005 Geochim. Cosmochim. Acta 70, 1148–1162. <https://doi.org/10.1016/j.gca.2005.11.002>  
 1006 Peckmann, J., Paul, J., Thiel, V., 1999. Bacterially mediated formation of diagenetic aragonite and  
 1007 native sulfur in Zechstein carbonates (Upper Permian, Central Germany). Sediment. Geol.  
 1008 126, 205–222. [https://doi.org/10.1016/S0037-0738\(99\)00041-X](https://doi.org/10.1016/S0037-0738(99)00041-X)

- 1009 Planel, D., Sercombe, J., Le Bescop, P., Adenot, F., Torrenti, J.-M., 2006. Long-term performance of  
1010 cement paste during combined calcium leaching–sulfate attack: kinetics and size effect. *Cem.*  
1011 *Concr. Res.* 36, 137–143. <https://doi.org/10.1016/j.cemconres.2004.07.039>
- 1012 Qian, Z., Tianwei, H., Mackey, H.R., van Loosdrecht, M.C.M., Guanghao, C., 2019. Recent advances in  
1013 dissimilatory sulfate reduction: From metabolic study to application. *Water Res.* 150, 162–  
1014 181. <https://doi.org/10.1016/j.watres.2018.11.018>
- 1015 Rafrafi, Y., Durban, N., Bertron, A., Albrecht, A., Robinet, J.-C., Erable, B., 2017. Use of a continuous-  
1016 flow bioreactor to evaluate nitrate reduction rate of *Halomonas desiderata* in cementitious  
1017 environment relevant to nuclear waste deep repository. *Biochem. Eng. J.* 125, 161–170.  
1018 <https://doi.org/10.1016/j.bej.2017.05.016>
- 1019 Rafrafi, Y., Ranaivomanana, H., Bertron, A., Albrecht, A., Erable, B., 2015. Surface and bacterial  
1020 reduction of nitrate at alkaline pH: Conditions comparable to a nuclear waste repository. *Int.*  
1021 *Biodeterior. Biodegrad.* 101, 12–22. <https://doi.org/10.1016/j.ibiod.2015.03.013>
- 1022 Ragu Nandhakumar, S., Rajeshkumar, S., Anand, R.S., Malya, V., Dua, K., Ezhilarasan, D., Lakshmi, T.,  
1023 2022. Chapter 20 - Biogenic metal sulfide nanoparticles synthesis and applications for  
1024 biomedical and environmental technology, in: Abd-Elsalam, K.A., Periakaruppan, R.,  
1025 Rajeshkumar, S. (Eds.), *Agri-Waste and Microbes for Production of Sustainable*  
1026 *Nanomaterials, Nanobiotechnology for Plant Protection*. Elsevier, pp. 495–506.  
1027 <https://doi.org/10.1016/B978-0-12-823575-1.00027-5>
- 1028 Ravot, G., Garcia, J., Magot, M., Ollivier, B., 2015. *Fusibacter*.  
1029 <https://doi.org/10.1002/9781118960608.gbm00723>
- 1030 Rizoulis, A., Steele, H.M., Morris, K., Lloyd, J.R., 2012. The potential impact of anaerobic microbial  
1031 metabolism during the geological disposal of intermediate-level waste. *Mineral. Mag.* 76,  
1032 3261–3270. <https://doi.org/10.1180/minmag.2012.076.8.39>
- 1033 Rout, S.P., Charles, C.J., Garratt, E.J., Laws, A.P., Gunn, J., Humphreys, P.N., 2015. Evidence of the  
1034 Generation of Isosaccharinic Acids and Their Subsequent Degradation by Local Microbial  
1035 Consortia within Hyper-Alkaline Contaminated Soils, with Relevance to Intermediate Level  
1036 Radioactive Waste Disposal. *Plos One* 10, e0119164.  
1037 <https://doi.org/10.1371/journal.pone.0119164>
- 1038 Sellier, A., Lacarrière, L., Gonnouni, M.E., Bourbon, X., 2011. Behavior of HPC nuclear waste disposal  
1039 structures in leaching environment. *Nucl. Eng. Des. - NUCL ENG DES* 241, 402–414.  
1040 <https://doi.org/10.1016/j.nucengdes.2010.11.002>
- 1041 Sorokin, D.Y., Banciu, H.L., Muyzer, G., 2015. Functional microbiology of soda lakes. *Curr. Opin.*  
1042 *Microbiol.* 25, 88–96. <https://doi.org/10.1016/j.mib.2015.05.004>
- 1043 Sorokin, D.Y., Berben, T., Melton, E.D., Overmars, L., Vavourakis, C.D., Muyzer, G., 2014. Microbial  
1044 diversity and biogeochemical cycling in soda lakes. *Extremophiles* 18, 791–809.  
1045 <https://doi.org/10.1007/s00792-014-0670-9>
- 1046 Sorokin, D.Y., Tourova, T.P., Panteleeva, A.N., Muyzer, G., 2012. *Desulfonatrobacter acidivorans*  
1047 *gen. nov., sp. nov.* and *Desulfobulbus alkaliphilus sp. nov.*, haloalkaliphilic heterotrophic  
1048 sulfate-reducing bacteria from soda lakes. *Int. J. Syst. Evol. Microbiol.* 62, 2107–2113.  
1049 <https://doi.org/10.1099/ijs.0.029777-0>
- 1050 Sousa, J.A.B., Plugge, C.M., Stams, A.J.M., Bijmans, M.F.M., 2015. Sulfate reduction in a hydrogen fed  
1051 bioreactor operated at haloalkaline conditions. *Water Res.* 68, 67–76.  
1052 <https://doi.org/10.1016/j.watres.2014.09.035>
- 1053 van Loon, L.R., Hummel, W., 1999. The Degradation of Strong Basic Anion Exchange Resins and  
1054 Mixed-Bed Ion-Exchange Resins: Effect of Degradation Products on Radionuclide Speciation.  
1055 *Nucl. Technol.* 128, 388–401. <https://doi.org/10.13182/NT99-A3039>
- 1056 vanGinkel, C.G., Vandenbroucke, K.L., Stroo, C.A., 1997. Biological removal of EDTA in conventional  
1057 activated-sludge plants operated under alkaline conditions. *Bioresour. Technol.* 59, 151–155.  
1058 [https://doi.org/10.1016/s0960-8524\(96\)00158-7](https://doi.org/10.1016/s0960-8524(96)00158-7)
- 1059 Vijay, K., Murmu, M., Deo, S.V., 2017. Bacteria based self healing concrete – A review. *Constr. Build.*  
1060 *Mater.* 152, 1008–1014. <https://doi.org/10.1016/j.conbuildmat.2017.07.040>

- 1061 Vinsot, A., Mettler, S., Wechner, S., 2008a. In situ characterization of the Callovo-Oxfordian pore  
1062 water composition. *Phys. Chem. Earth Parts ABC* 33, S75–S86.  
1063 <https://doi.org/10.1016/j.pce.2008.10.048>
- 1064 Vinsot, A., Mettler, S., Wechner, S., 2008b. In situ characterization of the Callovo-Oxfordian pore  
1065 water composition. *Phys. Chem. Earth Parts ABC* 33, S75–S86.  
1066 <https://doi.org/10.1016/j.pce.2008.10.048>
- 1067 Voegel, C., Bertron, A., Erable, B., 2016. Mechanisms of cementitious material deterioration in biogas  
1068 digester. *Sci. Total Environ.* 571, 892–901. <https://doi.org/10.1016/j.scitotenv.2016.07.072>
- 1069 Voegel, C., Bertron, A., Erable, B., 2015. Biodeterioration of cementitious materials in biogas digester.  
1070 *Matér. Tech.* 103, 202. <https://doi.org/10.1051/mattech/2015023>
- 1071 Voegel, C., Durban, N., Bertron, A., Landon, Y., Erable, B., 2019a. Evaluation of microbial proliferation  
1072 on cementitious materials exposed to biogas systems. *Environ. Technol.* 1–11.  
1073 <https://doi.org/10.1080/09593330.2019.1567610>
- 1074 Voegel, C., Giroudon, M., Bertron, A., Patapy, C., Matthieu, P.L., Verdier, T., Erable, B., 2019b.  
1075 Cementitious materials in biogas systems: Biodeterioration mechanisms and kinetics in CEM I  
1076 and CAC based materials. *Cem. Concr. Res.* 124, 105815.  
1077 <https://doi.org/10.1016/j.cemconres.2019.105815>
- 1078 Widdel, F., Pfennig, N., 1981. Studies on dissimilatory sulfate-reducing bacteria that decompose fatty  
1079 acids. I. Isolation of new sulfate-reducing bacteria enriched with acetate from saline  
1080 environments. Description of *Desulfobacter postgatei* gen. nov., sp. nov. *Arch. Microbiol.*  
1081 129, 395–400. <https://doi.org/10.1007/BF00406470>
- 1082 Zheng, Y., Xiao, Y., Yang, Z.-H., Wu, S., Xu, H.-J., Liang, F.-Y., Zhao, F., 2014. The bacterial communities  
1083 of bioelectrochemical systems associated with the sulfate removal under different pHs.  
1084 *Process Biochem.* 49, 1345–1351. <https://doi.org/10.1016/j.procbio.2014.04.019>  
1085